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ISOENZYME VARIATION AND SYSTEMATIC RELATIONSHIPS AMONG FOUR XEROPHYTES OF GENUS FESTUCA **FROM BULGARIA**

Angelov G.^{1*}, I. Bednarska²

¹Institute of Biodiversity & Ecosystem Research, Bulgarian Academy of Sciences, Acad. G. Bonchev Str., bl. 23, 1113 Sofia, Bulgaria, e-mail:gbangv@bio.bas.bg

²Institute of Ecology of the Carpathians NAS of Ukraine, Kozelnytska str., 4 Lviv 79026, Ukraine, e-mail: ibednarska@ukr.net

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Summary: The isoenzyme variation of Festuca hirtovaginata (Acht.) Markgr. Dann., F. thracica (Acht.) Markgr. Dann., F. hercegovinica Markgr. Dann. and F. oviniformis Vett. was examined by means of four molecular markers. Systematic relationships among the above mentioned taxa were assessed by calculating coefficient of divergence D. The mean values of coefficient D averaged over the four enzymes varied in a rather narrow range from 0.21 to 0.31 for all pair-wise comparison among the studied species of genus Festuca.. It could be assumed that the studied taxa are well differentiated and closely related entities within genus Festuca. The species F. oviniformis and F. hirtovaginata proved to be most closely related for the set of isoenzyme markers employed. Festuca thracica was relatively more distantly positioned within the studied group. As a whole, it could be concluded that the four studied species are more or less closely related within genus Festuca. The acknowledgement of F. hirtovaginata and F. thracica as closely related but different species and the treatment of F. hercegovinica as a distinct species seemed justified.

Keywords: Festuca, xerophytes, isoenzyme variation, systematic relationships.

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INTRODUCTION

The present study included four xerophytes of genus Festuca, namely, hirtovaginata, F. thracica F. F_{\cdot} hercegovinica and F. oviniformis. The species F. hercegovinica is a newly describedtaxon(Markgraff-Dannenberg, 1980) that grows on silicates. The species F. oviniformis grows on serpentinites. Festuca hirtovaginata and F. thracica have varied taxonomic history. Earlier F. thracica was treated as variety/subspecies of F. ovina or both taxa were considered as forms of F. duriuscula (Achtarov, 1953). Later

^{*}Corresponding author: gbangv@bio.bas.bg

Markgraff-Dannenberg (1980) changed the taxonomical status ascribed by Achtarov and ranked *F. hirtovaginata* and *F. thracica* as species.

Isoenzymes can used be for evaluating genetic differences and relationships systematic within taxonomically complicated plant groups. In the last two decades several isoenzyme studies of fescues (Aiken et al., 1993; Aiken et al., 1994; Aiken and Lefkovitch, 1995; Guldahl et al., 2001) were carried out to investigate species delimitation.

The purpose of the present study was to reveal the isoenzyme variation and systematic relationships among the above-mentioned four species of genus *Festuca*.

MATERIALS AND METHODS

The enzymes anodal peroxidase (PER), EC 1.11.1.7, acid phosphatase (ACP), EC 3.1.3.2, superoxide dismutase (SOD), EC 1.15.1.1 and amylase (AMY), EC 3.2.1.1, were analyzed individually in natural populations of the above listed taxa. Anodally migrating isoforms were resolved on 7.5% polyacrilamide slab gels as described by Davis (1964). The length of gels was 11 cm for AMY and SOD, 9 cm for ACP and 6 cm for anodal PER. The following staining recipes were used: AMY (Reisfeld et al. 1962), PER (Przybylska et al., 1982), ACP (Shaw and Prasad, 1970), SOD (Baur and Schorr, 1969).

Systematic relationships among the above mentioned taxa of genus *Festuca* were assessed by calculating coefficient of divergence D (Stuessy, 1990) according to the following formula:

$$D = \sqrt{\frac{1}{N}\sum_{i=1}^{N} (x_{ij} - x_{ik})^2}$$

where, N is the number of isoforms for each enzyme, x_{ij} and x_{ik} are the mean frequencies of i-th isoform in taxa j and k and presented graphically as a dendrogram using the non-weighted pair-group method with arithmetic mean (UPGMA) (Statistica 7.0).

RESULTS AND DISCUSSION

Anodal peroxidase

Totally thirteen isoforms of the enzyme from the studied species were electrophoretically resolved (Table 1). Most of the isoforms were shared by all species. Except for F. thracica, isoform 19 was monomorphically fixed throughout the studied group. Similarly, isoform 48 was monomorphic in all taxa, but was absent in F. hercegovinica. Isoform 40 invariant was in F_{\cdot} hirtovaginata and F. thracica and it was absent in F. hercegovinica and F. oviniformis. The values of coefficient D varied in a wide range (0.18-0.50). Festuca hirtovaginata and F. thracica proved to be most closely positioned (D=0.18), while the species pair hirtovaginata-hercegovinica was most distant (D=0.50).

Acid phosphatase

In total, twelve isoforms of ACP were detected in the examined species of genus *Festuca* (Table 2). Most isoforms were common for all examined species. Similarly, isoforms 7, 32, 39 occurred in all studied taxa, but were absent in *F. oviniformis*. The values of coefficient

Table 1. Mean isoform frequencies of anodal peroxidase in the studied populations of F. hercegovinica, F. ovinoformis, F. thracica and F. hirtovaginata.

Canor							Isoform						
sarado	5	10	11	15	17	19	22	23	25	40	43	48	52
F. hercegovinica	0.00	00.0	0.43	0.00	0.00	1.00	0.40	0.62	0.00	0.00	0.76	0.81	0.95
F. oviniformis	0.20	0.06	0.13	0.13	0.13	1.00	0.33	0.20	0.20	0.00	0.93	1.00	1.00
F. thracica	0.13	0.07	0.14	0.00	0.13	0.67	0.20	0.20	0.13	1.00	1.00	1.00	0.00
F. hirtovaginata	0.00	0.00	0.20	0.47	0.05	1.00	0.05	0.00	0.00	1.00	1.00	1.00	0.00

Table 2. Mean isoforand F. hirtovaginata.	m freque	ncies of a	acid phos	phatase ii	n the stuc	lied popu	lations of	F. herce,	govinica,	F. ovinof	ormis, F.	thracica
0						Isof	orm		1			
opecies	7	6	13	17	19	23	32	35	37	39	42	47
F. hercegovinica	0.10	0.47	0.95	0.24	0.52	0.86	0.14	0.81	0.05	0.15	0.57	0.35
F. oviniformis	0.00	0.14	0.21	0.75	0.43	0.77	0.00	0.37	0.34	0.00	0.13	0.87
F. thracica	0.43	0.77	0.77	0.14	0.14	0.28	0.72	0.61	0.15	0.68	0.45	0.22
F. hirtovaginata	0.72	0.55	0.32	0.38	0.45	0.48	0.58	0.34	0.09	0.35	0.65	0.18

srcegovinica, F. ovinoformis, F. thracica	
the studied populations of F . he	
iencies of acid phosphatase ir	
able 2. Mean isoform freque	nd F. hirtovaginata.

D ranged from 0.25 (*F. hirtovaginata* vs *F. thracica*) to 0.50 when the species pairs *F. oviniformis* / *F. thracica* were compared.

Superoxide dismutase

Seven isoforms of SOD were observed in the studied species of genus *Festuca* (Table 3). Monomorphicallyfixed isoforms 9 and 57 were common for the whole species group. Except for *F. thracica*, isoform 32 was monomorphic throughout the studied group. Similarly, isoform 74 was invariant in all taxa, but was absent in *F. oviniformis*. The values of coefficient D fluctuated from 0.07 (*F. hercegovinica* vs *F. thracica*) to 0. 31 when the species *F. oviniformis* and *F. hirtovaginata* were contrasted.

Amylase

Totally nine isoforms of the enzyme were electrophoretically resolved in the examined species (Table 4). Isoforms 19, 25 and 34 were shared by all studied species. Except for *F. oviniformis*, isoforms 3, 23 and 44 were also common for the whole group. Similarly, isoforms 12 and 27 were observed in all studied taxa, but were absent in *F. thracica*. The values of coefficient D varied in a comparatively wide range - from 0.16 (*F. hercegovinica* vs *F. thracica*) to 0.29 when *F. hercegovinica* and *F. oviniformis* were compared.

The mean values of coefficient D averaged over the four enzymes surveyed are shown in Table 5. The values varied in a rather narrow range - from 0.21 to 0.31 for all pair-wise comparisons. It could be assumed that the studied taxa are well differentiated and closely related entities within genus *Festuca*.

Systematic relationships among the above mentioned taxa are presented graphically as a dendrogram (Fig.1). The species F. oviniformis and F. hirtovaginata proved to be most closely related. Festuca thracica was relatively more distantly positioned within the studied group. As a whole, the four studied species are more or less closely related within genus Festuca. Morphologically and ecologically the studied species demonstrated close affinity. Nevertheless, the presented data showed some quantitative differences in their isoenzyme structure. Other closely related taxa of genus Festuca were also differentiated due to fixed or close to

Second				Isoform			
Species	9	25	32	38	48	57	74
F. hercegovinica	1.00	0.00	1.00	0.66	0.33	1.00	1.00
F. oviniformis	1.00	1.00	1.00	0.40	0.60	1.00	0.80
F. thracica	1.00	0.80	0.80	0.65	0.25	1.00	1.00
F. hirtovaginata	1.00	0.63	1.00	0.95	0.15	1.00	1.00

Table 3. Mean isoform frequencies of superoxide dismutase in the studied populations of *F. hercegovinica, F. ovinoformis, F. thracica* and *F. hirtovaginata.*

		44	0.42	0.00	0.27	0.38
		34	0.05	0.18	0.15	0.33
0		27	0.08	0.24	0.00	0.10
		25	0.22	0.08	0.34	0.26
 	Isoform	23	0.48	0.00	0.10	0.14
		19	0.32	0.06	0.42	0.52
		12	0.00	0.35	0.00	0.07
		10	0.00	0,26	0.08	0.14
aginata.		3	0.12	0.00	0.18	0.08
thracica and F. hirtov	Caoo:	opecies	F. hercegovinica	F. oviniformis	F. thracica	F. hirtovaginata

Table 4. Mean isoform frequencies of amylase in the studied populations of F. hercegovinica, F. ovinoformis, F.

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	Spacing		Coeffi	cient D	
	Species	1	2	3	4
1	F. hercegovinica	Х			
2	F. oviniformis	0.26	Х		
3	F. thracica	0.25	0.29	Х	
4	F. hirtovaginata	0.30	0.31	0.21	х

Table 5. Mean values of coefficient D for each pair-wise comparison among the studied species of genus *Festuca*.



Figure 1. Dendrogram of Cluster analysis for the four studied *Festuca* species based on D coefficient.

being fixed differences (Aiken et al., 1993; Aiken and Lefkovitch, 1995; Guldahl et al., 2001). In this sense, the acknowledgement by Markgraff-Dannenberg (1980) of *F. hirtovaginata*

and *F. thracica* as closely related but different species and the treatment of *F. hercegovinica* as a distinct species (Markgraff-Dannenberg, 1980) seems justified.

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