

## СПИСЪК НА ЗАБЕЛЯЗАНИ ЦИТИРАНИЯ В WoS и Scopus

на гл. ас. д-р Кирил Михайлов Мишев  
(без автоцитати и полуавтоцитати)

Брой цитирани публикации: <b>19</b>	Брой цитиращи източници: <b>285</b>
--	--

---

### 2004

---

1. Doltchinkova, V, **Georgieva, K**, Traytcheva, N, Slavov, Ch, **Mishev, K**. Melittin-induced changes in thylakoid membranes: particle electrophoresis and light scattering study. Biophysical Chemistry, 109, 3, Elsevier, 2004, ISSN:0301-4622, Web of Science, DOI:10.1016/j.bpc.2003.10.030, 387-397. SJR (Scopus):0.85, JCR-IF (Web of Science):2.102

Цитира се в:

1. Lee JW. Designer Transgenic Algae for Photobiological Production of Hydrogen from Water, Advanced Biofuels and Bioproducts, Springer New York, 371-404., @2013 [Линк](#)

---

### 2007

---

2. **Ananieva, K**, Ananiev, ED, **Mishev, K**, **Georgieva, K**, Malbeck, J, Kaminek, M, van Staden, J. Methyl jasmonate is a more effective senescence-promoting factor in Cucurbita pepo (zucchini) cotyledons when compared with darkness at the early stage senescence. Journal of Plant Physiology, 164, 9, Elsevier, 2007, ISSN:0176-1617, SCOPUS, DOI:doi:10.1016/j.jplph.2006.07.008, 1179-1187. JCR-IF (Web of Science):2.239

Цитира се в:

2. Jasad S, Galatro A, Villordo JJ, Puntarulo S, Simontacchi M. Role of nitric oxide in soybean cotyledon senescence, Plant science, 176, 5, 662-668., @2009 [Линк](#)
3. Chen Y, Pang Q, Dai S, Wang Y, Chen S, Yan X. Proteomic identification of differentially expressed proteins in Arabidopsis in response to methyl jasmonate, Journal of plant physiology, 168, 10, 995-1008., @2011 [Линк](#)
4. Kovacik J., Klejdus B., Stork F., Hedbavni J., Backor M. - Comparison of methyl jasmonate and cadmium effect on selected physiological parameters in Scenedesmus quadricauda (Chlorophyta, Chlorophyceae). Journal of Phycology, 47 (5), 1044-1049, 2011, @2011
5. Tierranegra-García N, Salinas-Soto P, Torres-Pacheco I, Ocampo-Velázquez RV, Rico-García E, Mendoza-Díaz SO, Feregrino-Pérez AA, Mercado-Luna A, Vargas-Hernandez M, Soto-Zarazúa GM, Guevara-González RG. Effect of foliar salicylic acid and methyl jasmonate applications on protection against pill-bugs in lettuce plants (Lactuca sativa), Phytoparasitica, 39, 2, 137-144., @2011 [Линк](#)
6. Zubo YO, Yamburenko MV, Kusnetsov VV, Börner T. Methyl jasmonate, gibberellic acid, and auxin affect transcription and transcript accumulation of chloroplast genes in barley, Journal of plant physiology, 168, 12, 1335-1344., @2011 [Линк](#)
7. Wang CL, Liang ZS, Li DR, Liu Y, Liu FH. Effects of growth-regulating substances and soluble sugar contents on accumulation of salvianolic acids in salvia miltiorrhiza bunge, Zhiwu Shengli Xuebao/Plant Physiology Journal, 48, 2, 181-188, @2012 [Линк](#)
8. Miyamoto K, Oka M, Uheda E, Ueda J. Changes in metabolism of cell wall polysaccharides in oat leaves during senescence: relevance to the senescence-promoting effect of methyl jasmonate, Acta physiologicae plantarum, 35, 9, 2675-2683., @2013 [Линк](#)

9. Sarwat M, Naqvi AR, Ahmad P, Ashraf M, Akram NA. Phytohormones and microRNAs as sensors and regulators of leaf senescence: assigning macro roles to small molecules, *Biotechnology advances*, 31, 8, 1153-1171., @2013 [Линк](#)
10. Yan Z, Chen J, Li X. Methyl jasmonate as modulator of Cd toxicity in *Capsicum frutescens* var. *fasciculatum* seedlings, *Ecotoxicology and environmental safety*, 98, 203-209., @2013 [Линк](#)
11. Horbowicz M, Kosson R, Sempruch C, Debski H, Koczkodaj D. Effect of methyl jasmonate vapors on level of anthocyanins, biogenic amines and decarboxylases activity in seedlings of chosen vegetable species, *Acta Scientiarum Polonorum. Hortorum Cultus*, 13, 1, 3-15., @2014 [Линк](#)
12. Hanaka A, Maksymiec W, Bednarek W. The effect of methyl jasmonate on selected physiological parameters of copper-treated *Phaseolus coccineus* plants, *Plant Growth Regulation*, 77, 2, 167-177, @2015 [Линк](#)
13. Kumari P, Reddy CRK, Jha B. Methyl Jasmonate-Induced Lipidomic and Biochemical Alterations in the Intertidal Macroalga *Gracilaria dura* (Gracilariaceae, Rhodophyta), *Plant and Cell Physiology*, 56, 10, 1877-1889, @2015 [Линк](#)
14. Dučaiová Z, Sajko M, Mihaličová S, Repčák M. Dynamics of accumulation of coumarin-related compounds in leaves of *Matricaria chamomilla* after methyl jasmonate elicitation, *Plant Growth Regulation*, 79, 1, 81-94, @2016 [Линк](#)
15. Wilmowicz, E., Kućko, A., Frankowski, K., Świdziński, M., Marciniak, K., & Kopcewicz, J. (2016). Methyl jasmonate-dependent senescence of cotyledons in *Ipomoea nil*. *Acta Physiologiae Plantarum*, 38(9), 222., @2016 [Линк](#)
16. Khan T, Abbasi BH, Khan MA. The interplay between light, plant growth regulators and elicitors on growth and secondary metabolism in cell cultures of *Fagonia indica*. *Journal of Photochemistry and Photobiology B: Biology*, 185, 153-160, @2018 [Линк](#)
17. Andi SA, Gholami M, Ford CM, Maskani F. The effect of light, phenylalanine and methyl jasmonate, alone or in combination, on growth and secondary metabolism in cell suspension cultures of *Vitis vinifera*. *Journal of Photochemistry and Photobiology B: Biology*, 199, 111625., @2019 [Линк](#)
18. Papazian S, Girdwood T, Wessels BA, Poelman EH, Dicke M, Moritz T, Albrechtsen BR. Leaf metabolic signatures induced by real and simulated herbivory in black mustard (*Brassica nigra*). *Metabolomics*. 2019; 15(10):130., @2019 [Линк](#)

---

## 2008

---

3. **Ananieva K**, Ananiev ED, **Mishev K**, **Georgieva K**, Tzvetkova N, Van Staden J. Changes in photosynthetic capacity and polypeptide patterns during natural senescence and rejuvenation of *Cucurbita pepo* L. (zucchini) cotyledons. *Plant Growth Regulation*, 54, 1, Springer, 2008, ISSN:0167-6903, Web of Science, DOI:10.1007/s10725-007-9223-x, 23-29. ISI IF:1.333

Цитира се в:

19. Jasad S, Galatro A, Villordo JJ, Puntarulo S, Simontacchi M. Role of nitric oxide in soybean cotyledon senescence, *Plant science*, 176, 5, 662-668., @2009 [Линк](#)
20. Jiang L, Kong XW, Cao SQ, Zhang RX. Hydrogen peroxide is involved in regulation of tobacco leaf senescence. *Journal of Molecular Cell Biology*, 2009, 42(1), 82-88, @2009
21. Du J, Li M, Kong D, Wang L, Lv Q, Wang J, Bao F, Gong Q, Xia J, He Y. Nitric oxide induces cotyledon senescence involving co-operation of the NES1/MAD1 and EIN2-associated ORE1 signalling pathways in *Arabidopsis*, *Journal of experimental botany*, 65 (14): 4051-4063, @2013 [Линк](#)
22. Humby PL, Snyder ECR, Durnford DG. Conditional senescence in *Chlamydomonas reinhardtii* (Chlorophyceae). *Journal of Phycology*, 2013, 49 (2): 389-400, @2013
23. Wei XD, Shi DW, Chen GX. Physiological, structural, and proteomic analysis of chloroplasts during natural senescence of Ginkgo leaves, *Plant Growth Regulation*, 69, 2, 191-201., @2013 [Линк](#)
24. Hejnák V, Hnilíčková H, Hnilíčka F. Effect of ontogeny, heterophylly and leaf position on the gas exchange of the hop plant, *Plant, Soil and Environment*, 60, 11, 525-30., @2014 [Линк](#)
25. Sun JL, Sui XL, Huang HY, Wang SH, Wei YX, Zhang ZX. Low Light Stress Down-Regulated Rubisco Gene Expression and Photosynthetic Capacity During Cucumber (*Cucumis sativus* L.) Leaf Development, *Journal of Integrative Agriculture*, 13, 5, 997-1007., @2014 [Линк](#)

26. Lambert, R., Cabello-Díaz, J. M., Quiles, F. A., & Piedras, P. (2016). Identification of nucleases related to nutrient mobilization in senescing cotyledons from French bean. *Acta Physiologiae Plantarum*, 38(11), 266., @2016 [Линк](#)
27. Wilnowicz, E., Kućko, A., Frankowski, K., Świdziński, M., Marciniak, K., & Kopcewicz, J. (2016). Methyl jasmonate-dependent senescence of cotyledons in *Ipomoea nil*. *Acta Physiologiae Plantarum*, 38(9), 222., @2016 [Линк](#)
28. Zhang Z, Sun Y, Li Y. "Plant rejuvenation: from phenotypes to mechanisms". *Plant Cell Reports*, 2020; 39: 1249–1262., @2020 [Линк](#)
29. Diaz-Baena M, Delgado-García E, Pineda M, Galvez-Valdivieso G, Piedras P. "S-Like Ribonuclease T2 Genes Are Induced during Mobilisation of Nutrients in Cotyledons from Common Bean". *Agronomy*, 2021, 11(3): 490., @2021 [Линк](#)

---

## 2009

---

4. **Mishev K**, Stefanov D, **Ananieva K**, Slavov Ch, Ananiev ED. Different effects of dark treatment on pigment composition and photosystem I and II activities in intact cotyledons and primary leaves of *Cucurbita pepo* (zucchini). *Plant Growth Regulation*, 58, 1, Springer, 2009, ISSN:0167-6903, Web of Science, DOI:10.1007/s10725-008-9352-x, 61-71. JCR-IF (Web of Science):1.53

Цитира се в:

30. Falqueto AR, Silva FS, Cassol D, Magalhães Júnior AM, Oliveira AC, Bacarin MA. Chlorophyll fluorescence in rice: probing of senescence driven changes of PSII activity on rice varieties differing in grain yield capacity, *Brazilian Journal of Plant Physiology*, 22, 1, 35-41, @2010 [Линк](#)
31. Chen C, Wang J, Zhao X. Leaf senescence induced by EGY1 defection was partially restored by glucose in *Arabidopsis thaliana*, *Botanical Studies*, 57, 5, @2016 [Линк](#)
32. Akpinar A, Cansev A, Isleyen M. "Effects of the lichen *Peltigera canina* on *Cucurbita pepo* spp. *pepo* grown in soil contaminated by DDTs". *Environmental Science and Pollution Research*, 2021, 28(12): 14576-85., @2021 [Линк](#)
33. Gutbrod P, Yang W, Grujicic GV, Peisker H, Gutbrod K, Du LF, Dörmann P. "Phytol derived from chlorophyll hydrolysis in plants is metabolized via phytenal". *Journal of Biological Chemistry*, 2021 Mar 11: 100530, DOI: 10.1016/j.jbc.2021.100530, @2021 [Линк](#)

---

## 2011

---

5. **Mishev K**, **Dimitrova A**, Ananiev ED. Darkness affects differentially the expression of plastid-encoded genes and delays the senescence-induced down-regulation of chloroplast transcription in cotyledons of *Cucurbita pepo* L. (Zucchini). *Z. Naturforsch. C*, 66c, 3-4, De Gruyter, 2011, ISSN:1865-7125, Web of Science, DOI:0939 – 5075/2011/0300 – 0159, 159-166. ISI IF:0.772

Цитира се в:

34. Yamburenko MV, Zubo YO, Vanková R, Kusnetsov VV, Kulaeva ON, Börner T. Abscisic acid represses the transcription of chloroplast genes, *Journal of experimental botany*, 64 (14): 4491-4502, @2013 [Линк](#)
  35. Yamburenko M V, Zubo Y O, Börner T. Abscisic acid affects transcription of chloroplast genes via protein phosphatase 2C-dependent activation of nuclear genes: repression by guanosine-3'-5'-bisdiphosphate and activation by sigma factor 5, *The Plant Journal*, 82, 6, 1030–1041, DOI: 10.1111/tpl.12876, @2015 [Линк](#)
  36. Zhang C, Zhu Q, Liu S, Gao P, Zhu Z, Wang X, Luan F. "The complete chloroplast genome sequence of the *Cucurbita pepo* L. (Cucurbitaceae)". *Mitochondrial DNA Part B*, 3(2):717-8., @2018 [Линк](#)
6. **Mishev K**, Ananiev ED, Humbeck K. Organ-specific effects of dark treatment on photosynthesis and the expression of photosynthesis-related genes. *Biologia Plantarum*, 55, 2, Springer, 2011, ISSN:1573-8264 (Online), 269-278. JCR-IF (Web of Science):1.974

Цитира се в:

37. Poór P, Borbély PG, Bódi N, Bagyánszki M, Görgényi Miklósné Tari I. "Effects of salicylic acid on photosynthetic activity and chloroplast morphology under light and prolonged darkness". *Photosynthetica*. 2019; 57:367-76., @2019 [Линк](#)

7. **Ananieva K**, Ananiev ED, **Doncheva S**, Stefanov D, **Mishev K**, Kaminek M, Motyka V, Dobrev P, Malbeck J. Local induction of senescence by darkness in Cucurbita pepo (zucchini) cotyledons or the primary leaf induces opposite effects in the adjacent illuminated organ. *Plant Growth Regulation*, 65, 3, Springer, 2011, ISSN:0167-6903, SCOPUS, DOI:10.1007/s10725-011-9616-8, 459-471. JCR-IF (Web of Science):1.604

Цитупа се е:

38. Wu HY, Tang HK, Liu LA, Shi L, Zhang WF, Jiang CD. "Local weak light induces the improvement of photosynthesis in adjacent illuminated leaves in maize seedlings". *Physiologia Plantarum*, 2021, 171(1): 125-36., @2021 [Линк](#)

---

## 2012

---

8. Irani NG, Di Rubbo S, Mylle E, Schneider-Pizon J, Van Den Begin J, Hnilikova J, Sisa M, Vilarrasa-Blasi J, Szatmari AM, Van Damme D, **Mishev K**, Codreanu M-C, Kohout L, Strnad M, Cano-Delgado AI, Friml J, Madder A, Russinova E. Fluorescent castasterone reveals BRI1 signaling from the plasma membrane. *Nature Chemical Biology*, 8, Nature Publishing Group, 2012, ISSN:1552-4450, Web of Science, DOI:10.1038/nchembio.958, 583-589. ISI IF:12.948

Цитупа се е:

39. Beck M, Zhou J, Faulkner C, MacLean D, Robatzek S. Spatio-temporal cellular dynamics of the Arabidopsis flagellin receptor reveal activation status-dependent endosomal sorting. *The Plant Cell*, 24, 10, 4205-4219., @2012 [Линк](#)

40. Šamajová O, Takáč T, von Wangenheim D, Stelzer E, Šamaj J. Update on methods and techniques to study endocytosis in plants. *Endocytosis in Plants*, Springer Berlin Heidelberg, 1-36., @2012 [Линк](#)

41. Bücherl CA, van Esse GW, Kruis A, Luchtenberg J, Westphal AH, Aker J, van Hoek A, Albrecht C, Borst JW, de Vries SC. Visualization of BRI1 and BAK1 (SERK3) membrane receptor heterooligomers during brassinosteroid signaling. *Plant physiology*, 162, 4, 1911-1925., @2013 [Линк](#)

42. Choi SW, Tamaki T, Ebine K, Uemura T, Ueda T, Nakano A. RABA members act in distinct steps of subcellular trafficking of the FLAGELLIN SENSING2 receptor. *The Plant Cell*, 25, 3, 1174-1187., @2013 [Линк](#)

43. Fridman Y, Savaldi-Goldstein S. Brassinosteroids in growth control: how, when and where. *Plant science*, 209, 24-31., @2013 [Линк](#)

44. Hao J, Yin Y, Fei SZ. Brassinosteroid signaling network: implications on yield and stress tolerance. *Plant cell reports*, 32, 7, 1017-1030., @2013 [Линк](#)

45. Jiang J, Zhang C, Wang X. Ligand perception, activation, and early signaling of plant steroid receptor brassinosteroid insensitive 1. *Journal of integrative plant biology*, 55, 12, 1198-1211., @2013 [Линк](#)

46. Oklešt J, Rárová L, Strnad M. Brassinosteroids and their Biological Activities. *Natural Products*, Springer Berlin Heidelberg, 3851-3871., @2013 [Линк](#)

47. Rasmussen A, Heugebaert T, Matthys C, Van Deun R, Boyer FD, Goormachtig S, Stevens C, Geelen D. A fluorescent alternative to the synthetic strigolactone GR24. *Molecular plant*, 6, 1, 100-112., @2013 [Линк](#)

48. Shani E, Weinstain R, Zhang Y, Castillejo C, Kaiserli E, Chory J, Tsien RY, Estelle M. Gibberellins accumulate in the elongating endodermal cells of Arabidopsis root. *Proceedings of the National Academy of Sciences*, 110, 12, 4834-4839., @2013 [Линк](#)

49. Sharma I, Kaur N, Saini S, Pati PK. Emerging Dynamics of Brassinosteroids Research. *Biotechnology: Prospects and Applications*, Springer India, 3-17., @2013 [Линк](#)

50. Spallek T, Beck M, Khaled SB, Salomon S, Bourdais G, Schellmann S, Robatzek S. ESCRT-I mediates FLS2 endosomal sorting and plant immunity. *PLOS Genetics*, 9(12), e1004035, @2013 [Линк](#)

51. Tian M, Xie Q. Non-26S Proteasome Proteolytic Role of Ubiquitin in Plant Endocytosis and Endosomal Trafficking. *Journal of integrative plant biology*, 55, 1, 54-63., @2013 [Линк](#)

52. Wang C, Yan X, Chen Q, Jiang N, Fu W, Ma B, Liu J, Li C, Bednarek SY, Pan J. Clathrin light chains regulate clathrin-mediated trafficking, auxin signaling, and development in Arabidopsis, *The Plant Cell*, 25, 2, 499-516., @2013 [Линк](#)
53. Wells DM, Laplace L, Bennett MJ, Vernoux T. Biosensors for phytohormone quantification: challenges, solutions, and opportunities, *Trends in plant science*, 18, 5, 244-249., @2013 [Линк](#)
54. Xin P, Yan J, Fan J, Chu J, Yan C. A dual role of boronate affinity in high-sensitivity detection of vicinal diol brassinosteroids from sub-gram plant tissues via UPLC-MS/MS, *Analyst*, 138, 5, 1342-1345., @2013 [Линк](#)
55. Yamaoka S, Shimono Y, Shirakawa M, Fukao Y, Kawase T, Hatsugai N, Tamura K, Shimada T, Hara-Nishimura I. Identification and dynamics of Arabidopsis adaptor protein-2 complex and its involvement in floral organ development, *The Plant Cell*, 25, 8, 2958-2969., @2013 [Линк](#)
56. Zhu JY, Sae-Seaw J, Wang ZY. Brassinosteroid signalling, *Development*, 140, 8, 1615-1620., @2013 [Линк](#)
57. Band LR, Wells DM, Fozard JA, Ghetiu T, French AP, Pound MP, Wilson MH, Yu L, Li W, Hijazi HI, Oh J. Systems analysis of auxin transport in the Arabidopsis root apex, *The Plant Cell*, 26, 3, 862-875., @2014 [Линк](#)
58. Bar M, Avni A. Endosomal trafficking and signaling in plant defense responses, *Current opinion in plant biology*, 22, 86-92., @2014 [Линк](#)
59. Barbosa IC, Zourelidou M, Willige BC, Weller B, Schwechheimer C. D6 PROTEIN KINASE activates auxin transport-dependent growth and PIN-FORMED phosphorylation at the plasma membrane, *Developmental cell*, 29, 6, 674-685., @2014 [Линк](#)
60. Belkhadir Y, Yang L, Hetzel J, Dangi JL, Chory J. The growth–defense pivot: crisis management in plants mediated by LRR-RK surface receptors, *Trends in biochemical sciences*, 39, 10, 447-456., @2014 [Линк](#)
61. Bitterlich M, Krügel U, Boldt-Burisch K, Franken P, Kühn C. The sucrose transporter SISUT2 from tomato interacts with brassinosteroid functioning and affects arbuscular mycorrhiza formation, *The Plant Journal*, 78, 5, 877-889., @2014 [Линк](#)
62. Dimopoulos S, Mayer CE, Rudolf F, Stelling J. Accurate cell segmentation in microscopy images using membrane patterns, *Bioinformatics*, 30, 18, 2644-2651., @2014 [Линк](#)
63. Fàbregas N, Caño-Delgado AI. Turning on the microscope turret: a new view for the study of brassinosteroid signaling in plant development, *Physiologia plantarum*, 151, 2, 172-183., @2014 [Линк](#)
64. Fonseca S, Rosado A, Vaughan-Hirsch J, Bishopp A, Chini A. Molecular locks and keys: the role of small molecules in phytohormone research, *Frontiers in plant science*, 5, 709., @2014 [Линк](#)
65. Hayashi KI, Nakamura S, Fukunaga S, Nishimura T, Jenness MK, Murphy AS, Motose H, Nozaki H, Furutani M, Aoyama T. Auxin transport sites are visualized in planta using fluorescent auxin analogs, *Proceedings of the National Academy of Sciences*, 111, 31, 11557-11562., @2014 [Линк](#)
66. Luschnig C, Vert G. The dynamics of plant plasma membrane proteins: PINs and beyond, *Development*, 141, 15, 2924-2938., @2014 [Линк](#)
67. Ma Q, Robert S. Auxin biology revealed by small molecules, *Physiologia plantarum*, 151, 1, 25-42., @2014 [Линк](#)
68. Rigal A, Ma Q, Robert S. Unraveling plant hormone signaling through the use of small molecules, *Frontiers in plant science*, 5, 373., @2014 [Линк](#)
69. Robatzek S. "Endocytosis: At the crossroads of pattern recognition immune receptors and pathogen effectors". *Applied Plant Cell Biology. Plant Cell Monographs*, Springer, Berlin, Heidelberg, 2014, 22: 273-297., @2014 [Линк](#)
70. Robinson DG, Pimpl P. Receptor-mediated transport of vacuolar proteins: a critical analysis and a new model, *Protoplasma*, 251, 1, 247-264., @2014 [Линк](#)
71. Shirakawa M, Ueda H, Shimada T, Kohchi T, Hara-Nishimura I. Myosin Cell Development Is Regulated by Endocytosis Machinery and PIN1 Polarity in Leaf Primordia of Arabidopsis thaliana, *The Plant Cell*, 26, 11, 4448-4461., @2014 [Линк](#)
72. Smith JM, Leslie ME, Robinson SJ, Korasick DA, Zhang T, Backues SK, Cornish PV, Koo AJ, Bednarek SY, Heese A. Loss of Arabidopsis thaliana Dynamin-Related Protein 2B reveals separation of innate immune signaling pathways, *PLoS pathogens*, 10, 12, e1004578., @2014 [Линк](#)

73. Smith JM, Salamango DJ, Leslie ME, Collins CA, Heese A. Sensitivity to Flg22 is modulated by ligand-induced degradation and de novo synthesis of the endogenous flagellin-receptor FLAGELLIN-SENSING2, *Plant physiology*, 164, 1, 440-454., @2014 [Линк](#)
74. Wang J, Lei Z, Wen Y, Mao G, Wu H, Xu H. A Novel Fluorescent Conjugate Applicable To Visualize the Translocation of Glucose-Fipronil, *Journal of agricultural and food chemistry*, 62, 35, 8791-8798., @2014 [Линк](#)
75. Worden N, Girke T, Drakakaki G. Endomembrane Dissection Using Chemically Induced Bioactive Clusters, *Plant Chemical Genomics, Methods in Molecular Biology*, Humana Press, 2014, 1056: 159-168., @2014 [Линк](#)
76. Baral A, Shruithi KS, Mathew MK. Vesicular trafficking and salinity responses in plants, *IUBMB life*, 67, 9, 677-686, @2015 [Линк](#)
77. Belkhadir Y, Jaillais Y. The molecular circuitry of brassinosteroid signaling, *New Phytologist*, 206, 2, 522-540, @2015 [Линк](#)
78. Ben Khaled S, Postma J, Robatzek S. A moving view: subcellular trafficking processes in pattern recognition receptor-triggered plant immunity, *Annual review of phytopathology*, 53, 379-402, @2015 [Линк](#)
79. Bozkurt TO, Belhaj K, Dagdas YF, Chaparro-Garcia A, Wu CH, Cano LM, Kamoun S. Rerouting of Plant Late Endocytic Trafficking Toward a Pathogen Interface, *Traffic*, 16, 2, 204-226, @2015 [Линк](#)
80. Cardona-López X, Cuyas L, Marín E, Rajulu C, Irigoyen ML, Gil E, Puga MI, Bligny R, Nussaume L, Geldner N, Paz-Ares J. ESCRT-III-Associated Protein ALIX Mediates High-Affinity Phosphate Transporter Trafficking to Maintain Phosphate Homeostasis in Arabidopsis, *The Plant Cell*, 27, 9, 2560-2581, @2015 [Линк](#)
81. Chaparro-Garcia A, Schwizer S, Sklenar J, Yoshida K, Petre B, Bos JI, Schornack S, Jones AM, Bozkurt TO, Kamoun S. Phytophthora infestans RXLR-WY Effector AVR3a Associates with Dynamin-Related Protein 2 Required for Endocytosis of the Plant Pattern Recognition Receptor FLS2, *PLoS one*, 10, 9, e0137071, @2015 [Линк](#)
82. Fan L, Li R, Pan J, Ding Z, Lin J. Endocytosis and its regulation in plants, *Trends in plant science*, 20, 6, 388-397, @2015 [Линк](#)
83. Jelínková A, Müller K, Fílová-Pařezová M, Petrášek J. NiGNL1a ARF-GEF acts in endocytosis in tobacco cells, *BMC plant biology*, 15, 1, 272, @2015 [Линк](#)
84. Khripach VA, Zhabinskii VN, Ermolovich YV. "Synthetic Aspects of Brassinosteroids". In: *Studies in Natural Products Chemistry 2015*, Vol. 44, pp. 309-352. Elsevier., @2015 [Линк](#)
85. Malachowska-Ugarte M, Sperduto C, Ermolovich YV, Sauchuk AL, Jurásek M, Litvinovskaya RP, Straltsova D, Smolich I, Zhabinskii VN, Drašar P, Demidchik V. Brassinosteroid-BODIPY conjugates: Design, synthesis, and properties, *Steroids*, 102, 53-59, @2015 [Линк](#)
86. Martins S, Dohmann EM, Cayrel A, Johnson A, Fischer W, Pojer F, Satiat-Jeunemaître B, Jaillais Y, Chory J, Geldner N, Vert G. Internalization and vacuolar targeting of the brassinosteroid hormone receptor BRI1 are regulated by ubiquitination, *Nature communications*, 6, 6151, @2015 [Линк](#)
87. Waadt R, Hsu PK, Schroeder JI. Abscisic acid and other plant hormones: Methods to visualize distribution and signaling, *BioEssays*, 37, 12, 1338-1349, @2015 [Линк](#)
88. Wang L, Li H, Lv X, Chen T, Li R, Xue Y, Jiang J, Jin B, Baluška F, Šamaj J, Wang X. Spatiotemporal dynamics of the BRI1 receptor and its regulation by membrane microdomains in living Arabidopsis cells, *Molecular plant*, 8, 9, 1334-1349, @2015 [Линк](#)
89. Yao TS, Zhu XF, Jung JH, Xuan YH. Qa-SNARE Protein SYP22 Negatively Regulates Brassinosteroid Signaling in the Dark, *Acta Biologica Cracoviensia s. Botanica*, 57, 2, 79-88, @2015 [Линк](#)
90. Chakrabarti A, Velusamy T, Tee CY, Jones DA. "A mutational analysis of the cytosolic domain of the tomato Cf-9 disease-resistance protein shows that membrane-proximal residues are important for Avr9-dependent necrosis". *Molecular plant pathology*. 2016; 17(4):565-76., @2016 [Линк](#)
91. Hatsugai N, Hillmer R, Yamaoka S, Hara-Nishimura I, Katagiri F. The  $\mu$  Subunit of Arabidopsis Adaptor Protein-2 Is Involved in Effector-Triggered Immunity Mediated by Membrane-Localized Resistance Proteins, *Molecular Plant-Microbe Interactions*, 29, 5, 345-351, @2016 [Линк](#)
92. Ho CM, Paciorek T, Abrash E, Bergmann DC. Modulators of stomatal lineage signal transduction alter membrane contact sites and reveal specialization among ERECTA kinases, *Developmental Cell*, 38, 4, 345-357, @2016 [Линк](#)



93. Ito E, Uemura T, Ueda T, Nakano A. Distribution of RAB5-positive multivesicular endosomes and the trans-Golgi network in root meristematic cells of *Arabidopsis thaliana*, *Plant Biotechnology*, 2016, 33(4): 281-286, @2016 [Линк](#)
94. Jaillais Y, Vert G. Brassinosteroid signaling and BRI1 dynamics went underground, *Current Opinion in Plant Biology*, 33, 92-100, @2016 [Линк](#)
95. Lace B, Prandi C. Shaping Small Bioactive Molecules to Untangle Their Biological Function: A Focus on Fluorescent Plant Hormones, *Molecular Plant*, 9, 8, 1099-1118, @2016 [Линк](#)
96. Mbengue M, Bourdais G, Gervasi F, Beck M, Zhou J, Spallek T, Bartels S, Boller T, Ueda T, Kuhn H, Robatzek S. Clathrin-dependent endocytosis is required for immunity mediated by pattern recognition receptor kinases, *Proceedings of the National Academy of Sciences*, 113, 39, 11034-11039, @2016 [Линк](#)
97. Nemhauser JL, Torii KU. Plant synthetic biology for molecular engineering of signalling and development, *Nature plants*, 2, 16010, @2016 [Линк](#)
98. Niehl A, Wyrtsch I, Boller T, Heinlein M. Double-stranded RNAs induce a pattern-triggered immune signaling pathway in plants, *New Phytologist*, 2016, 211(3): 1008-1019, @2016 [Линк](#)
99. Nishimura K, Matsunami E, Yoshida S, Kohata S, Yamauchi J, Jisaka M, Nagaya T, Yokota K, Nakagawa T. The tyrosine-sorting motif of the vacuolar sorting receptor VSR4 from *Arabidopsis thaliana*, which is involved in the interaction between VSR4 and AP1M2,  $\mu$ 1-adaptin type 2 of clathrin adaptor complex 1 subunits, participates in the post-Golgi sorting of VSR4, *Bioscience, biotechnology, and biochemistry*, 80, 4, 694-705, @2016 [Линк](#)
100. Paez Valencia J, Goodman K, Otegui MS. Endocytosis and Endosomal Trafficking in Plants, *Annual review of plant biology*, 67, 309-335, @2016 [Линк](#)
101. Yoshinari A, Fujimoto M, Ueda T, Inada N, Naito S, Takano J. DRP1-dependent Endocytosis is Essential for Polar Localization and Boron-induced Degradation of the Borate Transporter BOR1 in *Arabidopsis thaliana*, *Plant and Cell Physiology*, 2016, 57(9): 1985-2000, @2016 [Линк](#)
102. Yu H, Huang T. Molecular Mechanisms of Floral Boundary Formation in *Arabidopsis*, *International journal of molecular sciences*, 17, 3, 317, @2016 [Линк](#)
103. Žárský V. Clathrin in plant defense signaling and execution, *Proceedings of the National Academy of Sciences*, 113, 39, 10745-10747., @2016 [Линк](#)
104. Bücherl CA, Jarsch IK, Schudoma C, Segonzac C, Mbengue M, Robatzek S, MacLean D, Ott T, Zipfel C. "Plant immune and growth receptors share common signalling components but localise to distinct plasma membrane nanodomains". *Elife*. 6, e25114, @2017 [Линк](#)
105. Collins CA, Leslie ME, Peck SC, Heese A. "Simplified Enrichment of Plasma Membrane Proteins from *Arabidopsis thaliana* Seedlings Using Differential Centrifugation and Brij-58 Treatment". *Brassinosteroids: Methods and Protocols*. 1564: 155-168, @2017 [Линк](#)
106. Hutten SJ, Hamers DS, Den Toorn MA, Van Esse W, Nolles A, Bücherl CA, De Vries SC, Hohlbein J, Borst JW. "Visualization of BRI1 and SERK3/BAK1 Nanoclusters in *Arabidopsis* Roots". *PLoS one*. 12, 1, e0169905, @2017 [Линк](#)
107. Jelenska J, Davern SM, Standaert RF, Mirzadeh S, Greenberg JT. "Flagellin peptide flg22 gains access to long-distance trafficking in *Arabidopsis* via its receptor, FLS2". *Journal of Experimental Botany*. 68, 7, 1769-1783, @2017 [Линк](#)
108. Samodelov SL, Zurbriggen MD. "Quantitatively Understanding Plant Signaling: Novel Theoretical-Experimental Approaches". *Trends in plant science*. 22, 8, 685-704, @2017 [Линк](#)
109. Schayek H, Shani E, Weinstain R. Highlighting Gibberellins Accumulation Sites in *Arabidopsis thaliana* Root Using Fluorescently Labeled Gibberellins, *Plant Hormones: Methods and Protocols*, 1497: 91-97, @2017 [Линк](#)
110. Song X, Guo H, Zhang G, Wu Y, Wang G, Chen X, Fang R. "OsPRA2 fine-tunes rice brassinosteroid receptor". *Plant Signaling & Behavior*. 12, 3, e1257455, @2017 [Линк](#)
111. Wolf S. "Plant cell wall signalling and receptor-like kinases". *Biochemical Journal*. 474 (4): 471-492, @2017 [Линк](#)
112. Yamagami A, Saito C, Nakazawa M, Fujioka S, Uemura T, Matsui M, Sakuta M, Shinozaki K, Osada H, Nakano A, Asami T. "Evolutionarily conserved BIL4 suppresses the degradation of brassinosteroid receptor BRI1 and regulates cell elongation". *Scientific reports*. 7, 1, 5739, @2017 [Линк](#)
113. Cui Y, Li X, Yu M, Li R, Fan L, Zhu Y, Lin J. "Sterols regulate endocytic pathways during flg22-induced defense responses in *Arabidopsis*". *Development*. 145(19):dev165688., @2018 [Линк](#)

114. Geisler M. "Seeing is better than believing: visualization of membrane transport in plants". *Current opinion in plant biology*. 46:104-12., @2018 [Линк](#)
115. Gruszka D. "Crosstalk of the Brassinosteroid Signalosome with Phytohormonal and Stress Signaling Components Maintains a Balance between the Processes of Growth and Stress Tolerance". *International journal of molecular sciences*. 19(9):2675., @2018 [Линк](#)
116. Jiang K, Asami T. "Chemical regulators of plant hormones and their applications in basic research and agriculture". *Bioscience, biotechnology, and biochemistry*. 82(8):1265-1300., @2018 [Линк](#)
117. Kinoshita T, McCourt P, Asami T, Torii KU. "Plant Chemical Biology". *Plant and Cell Physiology*, 59(8): 1483–1486., @2018 [Линк](#)
118. Kubiasová K, Mik V, Nisler J, Hönig M, Husičková A, Spíchal L, Pěkná Z, Šamajová O, Doležal K, Plíhal O, Benková E. "Design, synthesis and perception of fluorescently labeled isoprenoid cytokinins". *Phytochemistry*. 150:1-11., @2018 [Линк](#)
119. Liu C, Shen W, Yang C, Zeng L, Gao C. "Knowns and unknowns of plasma membrane protein degradation in plants". *Plant science*. Vol. 272: 55-61., @2018 [Линк](#)
120. Liu Q, Vain T, Viotti C, Doyle SM, Tarkowská D, Novák O, Zipfel C, Sitbon F, Robert S, Hofius D. "Vacuole Integrity Maintained by DUF300 Proteins Is Required for Brassinosteroid Signaling Regulation". *Molecular Plant*. 11(4):553-67., @2018 [Линк](#)
121. Lv M, Li M, Chen W, Wang Y, Sun C, Yin H, He K, Li J. "Thermal-enhanced bri1-301 instability reveals a plasma membrane protein quality control system in plants". *Frontiers in Plant Science*. 9: 1620., @2018 [Линк](#)
122. Peng Y, Chen L, Li S, Zhang Y, Xu R, Liu Z, Liu W, Kong J, Huang X, Wang Y, Cheng B. "BRI1 and BAK1 interact with G proteins and regulate sugar-responsive growth and development in Arabidopsis". *Nature communications*. 9(1):1522., @2018 [Линк](#)
123. Raboanatahiry N, Chao H, Dalin H, Pu S, Yan W, Yu L, Wang B, Li M. "QTL Alignment for Seed Yield and Yield Related Traits in Brassica napus". *Frontiers in plant science*. 9: 1127., @2018 [Линк](#)
124. Reynolds GD, Wang C, Pan J, Bednarek SY. "Inroads into internalization: five years of endocytic exploration". *Plant physiology*. 176(1):208-18., @2018 [Линк](#)
125. Ruan Y, Halat LS, Khan D, Jancowski S, Ambrose C, Belmonte MF, Wasteneys GO. "The Microtubule-associated protein CLASP sustains cell proliferation through a brassinosteroid signaling negative feedback loop". *Current Biology*. 28(17):2718-29., @2018 [Линк](#)
126. Wang L, Zhang Y, Wang D, Wang M, Wang Y, Feng J. "Mitochondrial Signs and Subcellular Imaging Imply the Antifungal Mechanism of Carabrone against *Gaeumannomyces graminis* var. *tritici*". *Journal of agricultural and food chemistry*. 2018, 66(1): 81-90, @2018 [Линк](#)
127. Yamagami A, Chieko S, Sakuta M, Shinozaki K, Osada H, Nakano A, Asami T, Nakano T. "Brassinosteroids regulate vacuolar morphology in root meristem cells of *Arabidopsis thaliana*". *Plant signaling & behavior*. 13(4): e1417722., @2018 [Линк](#)
128. Yamamoto M, Nishio T, Nasrallah JB. "Activation of Self-Incompatibility Signaling in Transgenic *Arabidopsis thaliana* Is Independent of AP2-Based Clathrin-Mediated Endocytosis". *G3: Genes, Genomes, Genetics*. 8(7):2231-9., @2018 [Линк](#)
129. Zhao C, Zayed O, Yu Z, Jiang W, Zhu P, Hsu CC, Zhang L, Tao WA, Lozano-Durán R, Zhu JK. "Leucine-rich repeat extensin proteins regulate plant salt tolerance in *Arabidopsis*". *Proceedings of the National Academy of Sciences*. 115(51):13123-8., @2018 [Линк](#)
130. Aubry E, Dinant S, Vilaine F, Bellini C, Le Hir R. "Lateral transport of organic and inorganic solutes". *Plants*. 2019; 8(1):20., @2019 [Линк](#)
131. Dragwidge JM, Scholl S, Schumacher K, Gendall AR. "NHX-type Na<sup>+</sup> (K<sup>+</sup>)/H<sup>+</sup> antiporters are required for TGN/EE trafficking and endosomal ion homeostasis in *Arabidopsis thaliana*". *J Cell Sci*. 2019; 132(7):jcs226472., @2019 [Линк](#)
132. Ekanayake G, LaMontagne ED, Heese A. "Never Walk Alone: Clathrin-Coated Vesicle (CCV) Components in Plant Immunity". *Annual Review of Phytopathology*. 2019; 57:387-409., @2019 [Линк](#)
133. Gao J, Chaudhary A, Vaddepalli P, Nagel MK, Isono E, Schneitz K. "The *Arabidopsis* receptor kinase STRUBBELIG undergoes clathrin-dependent endocytosis". *Journal of Experimental Botany*. 2019 Apr 25., @2019 [Линк](#)
134. Hurski AL, Kukel AG, Liubina AI, Baradzenka AG, Straltsova D, Demidchik V, Drašar P, Zhabinskii VN, Khripach VA. "Regio- and stereoselective C–H functionalization of brassinosteroids". *Steroids*. 2019 Jun 1;146:92-8., @2019 [Линк](#)



135. Kretynin SV, Kolesnikov YS, Derevyanchuk MV, Kalachova TA, Blume YB, Khripach VA, Kravets VS. "Brassinosteroids application induces phosphatidic acid production and modify antioxidant enzymes activity in tobacco in calcium-dependent manner". *Steroids*. 2019 Jul 8:108444., @2019 [Линк](#)
  136. Niehl A, Heinlein M. "Perception of double-stranded RNA in plant antiviral immunity". *Molecular Plant Pathology*. 2019 Apr 3., @2019 [Линк](#)
  137. Rodriguez-Furlan C, Minina EA, Hicks GR. "Remove, Recycle, Degrade: Regulating Plasma Membrane Protein Accumulation". *The Plant Cell*. 2019; 31(12):2833-54., @2019 [Линк](#)
  138. Rozhon W, Akter S, Fernandez A, Poppenberger B. "Inhibitors of Brassinosteroid Biosynthesis and Signal Transduction". *Molecules*. 2019; 24(23):4372., @2019 [Линк](#)
  139. Verna C, Ravichandran SJ, Sawchuk MG, Linh NM, Scarpella E. "Coordination of tissue cell polarity by auxin transport and signaling". *Elife*. 2019 ;8., @2019 [Линк](#)
  140. Xue S, Zou J, Liu Y, Wang M, Zhang C, Le J. "Involvement of BIG5 and BIG3 in BRI1 Trafficking Reveals Diverse Functions of BIG-subfamily ARF-GEFs in Plant Growth and Gravitropism". *International Journal of Molecular Sciences*. 2019; 20(9):2339., @2019 [Линк](#)
  141. Zhang L, Liu Y, Zhu XF, Jung JH, Sun Q, Li TY, Chen LJ, Duan YX, Xuan YH. "SYP22 and VAMP727 regulate BRI1 plasma membrane targeting to control plant growth in Arabidopsis". *New Phytologist*. 2019 Feb 25., @2019 [Линк](#)
  142. Chen C, Yuan M, Song J, Liu Y, Xia Z, Yuan Y, Wang W, Xie Q, Guan X, Chen Q, Chen J. "Genome-wide identification and testing of superior reference genes for transcript normalization during analyses of flesh development in Asian pear cultivars". *Scientia Horticulturae*, 2020 Sep 20; 271:109459., @2020 [Линк](#)
  143. Hou B, Shen Y. "A Clathrin-Related Protein, SCD2/RRP1, Participates in Abscisic Acid Signaling in Arabidopsis". *Frontiers in Plant Science*, 2020 Jun 18; 11:892., @2020 [Линк](#)
  144. Mao J, Li J. "Regulation of Three Key Kinases of Brassinosteroid Signaling Pathway". *International Journal of Molecular Sciences*, 2020 Jan; 21(12):4340., @2020 [Линк](#)
  145. Wolf S. "Deviating from the Beaten Track: New Twists in Brassinosteroid Receptor Function". *International Journal of Molecular Sciences*, 2020 Jan; 21(5):1561., @2020 [Линк](#)
  146. Yu M, Li R, Cui Y, Chen W, Li B, Zhang X, Bu Y, Cao Y, Xing J, Jewaria PK, Li X. "The RALF1-FERONIA interaction modulates endocytosis to mediate control of root growth in Arabidopsis". *Development*, 2020 Jul 1; 147(13)., @2020 [Линк](#)
  147. Ekanayake G, Smith JM, Jones KB, Stiers HM, Robinson SJ, LaMontagne ED, Kostos PH, Cornish PV, Bednarek SY, Heese A. "DYNAMIN-RELATED PROTEIN DRP1A functions with DRP2B in plant growth, flg22-immune responses, and endocytosis". *Plant Physiology*, 2021 Feb 3, DOI: 10.1093/plphys/kiab024, @2021 [Линк](#)
  148. Isoda R, Yoshinari A, Ishikawa Y, Sadoine M, Simon R, Frommer WB, Nakamura M. "Sensors for the quantification, localization and analysis of the dynamics of plant hormones". *The Plant Journal*, 2021, 105(2): 542-57., @2021 [Линк](#)
  149. Starodubtseva A, Kalachova T, Iakovenko O, Stoudková V, Zhabinskii V, Khripach V, Ruelland E, Martinec J, Burketová L, Kravets V. "BODIPY Conjugate of Epibrassinolide as a Novel Biologically Active Probe for In Vivo Imaging". *International Journal of Molecular Sciences*, 2021, 22(7): 3599., @2021 [Линк](#)
  150. Turek I, Irving H. "Moonlighting proteins shine new light on molecular signaling niches". *International Journal of Molecular Sciences*, 2021, 22(3): 1367., @2021 [Линк](#)
9. **Ivanova A, Ananieva K, Mishev K, Ananiev ED.** Lipid composition in leaves and cotyledons of Cucurbita pepo L. (zucchini) during natural and induced senescence. *Genetics and Plant Physiology*, 2, 1-2, Prof. Marin Drinov Pub House of BAS, 2012, ISSN:1314-5770, EBSCO Publishing Inc., 98-106. ISI IF:0.27

Цитира се в:

151. Iosypenko OO, Kyslychenko VS, Omelchenko ZI, Burlaka IS. "Fatty acid composition of vegetable marrows and zucchini leaves". *Pharmacia*, 2019, 66 (4): 201-207, @2019 [Линк](#)
152. Stabler D, Al-Esawy M, Chennells JA, Perri G, Robinson A, Wright GA. "Regulation of dietary intake of protein and lipid by nurse-age adult worker honeybees". *Journal of Experimental Biology*, 2021 Feb 1; 224: jeb230615, DOI: 10.1242/jeb.230615, @2021 [Линк](#)

10. **Mishev K**, Dejonghe W, Russinova E. Small molecules for dissecting endomembrane trafficking: a cross-systems view. *Chemistry and Biology*, 20, 4, Cell Press, 2013, ISSN:1074-5521, Web of Science, 475-486. ISI IF:6.586

Цитира се в:

153. Ivanov AI. "Pharmacological Inhibitors of Exocytosis and Endocytosis: Novel Bullets for Old Targets". In: *Exocytosis and Endocytosis*, Springer New York, 2014, 1174: 3-18, @2014 [Линк](#)
154. Rubilar-Hernández C, Hicks GR, Norambuena L. "Chemical Genomics Screening for Biomodulators of Endomembrane System Trafficking". *Plant Endosomes*, Springer New York, 2014, 1209: 251-264, @2014 [Линк](#)
155. Doyle SM, Vain T, Robert S. "Small molecules unravel complex interplay between auxin biology and endomembrane trafficking". *Journal of Experimental Botany*, 2015, 66 (16): 4971-4982, @2015 [Линк](#)
156. Wu S, Gallagher KL. "Techniques for assessing the effects of pharmacological inhibitors on intercellular protein movement". *PLASMODESMATA: METHODS AND PROTOCOLS*, Book Series: Methods in Molecular Biology, Springer New York, 1217: 245-258, @2015 [Линк](#)
157. Baral A, Bhalerao RP. "Exploring exocytosis using chemical genomics". *Proceedings of the National Academy of Sciences of U S A*, 113 (1): 14-16, @2016 [Линк](#)
158. Zheng W, Zheng H, Zhao X, Zhang Y, Xie Q, Lin X, Chen A, Yu W, Lu G, Shim WB, Zhou J. "Retrograde trafficking from the endosome to the trans-Golgi network mediated by the retromer is required for fungal development and pathogenicity in *Fusarium graminearum*". *New Phytologist*, 2016, 210 (4): 1327-1343, @2016 [Линк](#)
159. Benabdi S, Peurois F, Nawrotek A, Chikireddy J, Cañeque T, Yamori T, Shiina I, Ohashi Y, Dan S, Rodriguez R, Cherfils J. "Family-wide Analysis of the Inhibition of Arf Guanine Nucleotide Exchange Factors with Small Molecules: Evidence of Unique Inhibitory Profiles". *Biochemistry*. 56, 38, 5125-5133, @2017 [Линк](#)
160. Jung JY, Lee DW, Ryu SB, Hwang I, Schachtman DP. "SCYL2 genes are involved in clathrin-mediated vesicle trafficking and essential for plant growth". *Plant Physiology*, 175 (1): 194-204, @2017 [Линк](#)
161. Alborzinia H, Ignashkova TI, Dejure FR, Gendarme M, Theobald J, Wöfl S, Lindemann RK, Reiling JH. "Golgi stress mediates redox imbalance and ferroptosis in human cells". *Communications biology*. 1(1):210., @2018 [Линк](#)
162. Bridges R.J., Bradbury N.A. "Cystic Fibrosis, Cystic Fibrosis Transmembrane Conductance Regulator and Drugs: Insights from Cellular Trafficking". In: Ulloa-Aguirre A., Tao YX. (eds) *Targeting Trafficking in Drug Development*. Handbook of Experimental Pharmacology, vol 245. Springer, Cham, @2018 [Линк](#)
163. Bruinsma S, James DJ, Serrano MQ, Esquibel J, Woo SS, Kielar-Grevstad E, Crummy E, Qurashi R, Kowalchuk JA, Martin TF. "Small molecules that inhibit the late stage of Munc13-4-dependent secretory granule exocytosis in mast cells". *Journal of Biological Chemistry*. 293(21):8217-29., @2018 [Линк](#)
164. Komatsu R, Yamaguchi T, Kobayashi N, Ozeki Y, Sakurai K. "Synthesis of alkyne-tagged and biotin-tagged Sortin1 as novel photoaffinity probes". *Bioorganic & medicinal chemistry letters*. 28(9):1562-5., @2018 [Линк](#)
165. Wu G, Cui X, Chen H, Renaud JB, Yu K, Chen X, Wang A. "Dynamin-like proteins of endocytosis in plants are coopted by potyviruses to enhance virus infection". *Journal of virology*. 92(23):e01320-18., @2018 [Линк](#)
166. Bhattacharyya A, Gupta A, Kuppasamy L, Mani S, Shukla A, Srivas M, Thattai M. "A formal methods approach to predicting new features of the eukaryotic vesicle traffic system". *Acta Informatica*. 2019: 1-37., @2019 [Линк](#)
167. Boncompain G, Gareil N, Tessier S, Lescure A, Jones TR, Kepp O, Kroemer G, Del Nery E, Perez F. "BML-265 and Tyrphostin AG1478 disperse the Golgi apparatus and abolish protein transport in human cells". *Frontiers in Cell and Developmental Biology*. 2019; 7:232., @2019 [Линк](#)
168. Demuyser L, Van Dyck K, Timmermans B, Van Dijck P. "Inhibition of vesicular transport influences fungal susceptibility to fluconazole". *Antimicrobial agents and chemotherapy*. 2019; 63(5):e01998-18., @2019 [Линк](#)

169. Huang L, Li X, Zhang C. "Progress in using chemical biology as a tool to uncover novel regulators of plant endomembrane trafficking". *Current Opinion in Plant Biology*. 2019; 52:106-13., @2019 [Линк](#)
170. Loebel C, Mauck RL, Burdick JA. "Local nascent protein deposition and remodelling guide mesenchymal stromal cell mechanosensing and fate in three-dimensional hydrogels". *Nature materials*. 2019 Mar 18:1., @2019 [Линк](#)
171. Iswanto AB, Shon JC, Liu KH, Vu MH, Kumar R, Kim JY. "Sphingolipids Modulate Secretion of Glycosylphosphatidylinositol-Anchored Plasmodesmata Proteins and Callose Deposition". *Plant Physiology*, 2020; 184(1):407-20., @2020 [Линк](#)
172. Lešková A, Labajová M, Krausko M, Zahradníková A, Baluška F, Mičieta K, Turňa J, Jásik J. "Endosidin 2 accelerates PIN2 endocytosis and disturbs intracellular trafficking of PIN2, PIN3, and PIN4 but not of SYT1". *PLoS one*, 2020; 15(8):e0237448., @2020 [Линк](#)

---

## 2014

---

11. Dejonghe W, **Mishev K**, Russinova E. The brassinosteroid chemical toolbox. *Current Opinion in Plant Biology*, 22, Elsevier, 2014, ISSN:1369-5266, Web of Science, DOI:10.1016/j.pbi.2014.09.002, 48-55. ISI IF:7.848

Цитира се е:

173. Flasiński M, Bartosik M, Kowal S, Broniatowski M, Wydro P. "Characteristics of the influence of auxins on physicochemical properties of membrane phospholipids in monolayers at the air/aqueous solution interface". *Colloids and Surfaces B: Biointerfaces*, 2015, 136: 1131-1138, @2015 [Линк](#)
174. Singh AP, Savaldi-Goldstein S. "Growth control: brassinosteroid activity gets context". *Journal of Experimental Botany*, 66 (4): 1123-1132, @2015 [Линк](#)
175. Flasiński M, Swiechowicz P. "Phytohormones Behavior in the Model Environment of Plant and Human Lipid Membranes". *The Journal of Physical Chemistry B*, 2017, 121 (25): 6175-6183, @2017 [Линк](#)
176. Sakaguchi J, Watanabe Y. "Light perception in aerial tissues enhances DWF4 accumulation in root tips and induces root growth". *Scientific Reports*. 7, 1808, @2017 [Линк](#)
177. Wang X, Teng Y, Zhang N, Christie P, Li Z, Luo Y, Wang J. "Rhizobial symbiosis alleviates polychlorinated biphenyls-induced systematic oxidative stress via brassinosteroids signaling in alfalfa". *Science of The Total Environment*. 592, 68-77, @2017 [Линк](#)
178. Li QF, Lu J, Yu JW, Zhang CQ, He JX, Liu QQ. "The brassinosteroid-regulated transcription factors BZR1/BES1 function as a coordinator in multesignal-regulated plant growth". *Biochimica et Biophysica Acta (BBA)-Gene Regulatory Mechanisms*, Vol. 1861 (6): 561-571., @2018 [Линк](#)
179. Kerchev P, van der Meer T, Sujeeth N, Verlee A, Stevens CV, Van Breusegem F, Gechev T. "Molecular priming as an approach to induce tolerance against abiotic and oxidative stresses in crop plants". *Biotechnology Advances*. 2019: 107503., @2019 [Линк](#)
180. Ono A, Sato A, Fujimoto KJ, Matsuo H, Yanai T, Kinoshita T, Nakamichi N. "3, 4-Dibromo-7-Azaindole Modulates Arabidopsis Circadian Clock by Inhibiting Casein Kinase 1 Activity". *Plant and Cell Physiology*. 2019; 60(11):2360-8., @2019 [Линк](#)
181. Rozhon W, Akter S, Fernandez A, Poppenberger B. "Inhibitors of Brassinosteroid Biosynthesis and Signal Transduction". *Molecules*. 2019; 24(23):4372., @2019 [Линк](#)

---

## 2016

---

12. Dejonghe, W, Kuenen S, Mylle E, Vasileva M, Keech O, Viotti C, Swerts J, Fendrych M, Ortiz-Morea FA, **Mishev K**, Delang S, Scholl S, Zarza X, Heilmann M, Kourelis J, Kasprovcz J, Nguyen le SL, Drozdzecki A, Van Houtte I, Szatmári AM, Majda M, Baisa G, Bednarek SY, Robert S, Audenaert D, Testerink C, Munnik T, Van Damme D, Heilmann I, Schumacher K, Winne J, Friml J, Verstreken P, Russinova E. Mitochondrial uncouplers inhibit clathrin-mediated endocytosis largely through cytoplasmic acidification. *Nature Communications*, 7, 11710, Springer Nature Publishing AG, 2016, ISSN:2041-1723, Web of Science, DOI:10.1038/ncomms11710, ISI IF:12.124

Цитира се е:

182. Bertone NI, Groisman AI, Mazzone GL, Cano R, Tabares L, Uchitel OD. "Carbonic anhydrase inhibitor acetazolamide shifts synaptic vesicle recycling to a fast mode at the mouse neuromuscular junction". *Synapse*. 71, 12, @2017 [Линк](#)
183. Bueno-Junior LS, Ruggiero RN, Rossignoli MT, Del Bel EA, Leite JP, Uchitel OD. "Acetazolamide potentiates the afferent drive to prefrontal cortex in vivo". *Physiological reports*. 5, 1, e13066, @2017 [Линк](#)
184. Gao W, Shi P, Chen X, Zhang L, Liu J, Fan X, Luo X. "Clathrin-mediated integrin  $\alpha\text{IIb}\beta\text{3}$  trafficking controls platelet spreading". *Platelets*. 1, 1-2, @2017 [Линк](#)
185. Murray JW, Yin D, Wolkoff AW. "Reduction of organelle motility by removal of potassium and other solutes". *PloS one*. 12, 9, e0184898, @2017 [Линк](#)
186. Norambuena L, Tejos R. "Chemical Genetic Dissection of Membrane Trafficking". *Annual Review of Plant Biology*. 68, 197-224, @2017 [Линк](#)
187. Underwood W, Ryan A, Somerville SC. "An Arabidopsis Lipid Flippase Is Required for Timely Recruitment of Defenses to the Host-Pathogen Interface at the Plant Cell Surface". *Molecular Plant*. 10, 6, 805-820, @2017 [Линк](#)
188. Childress ES, Alexopoulos SJ, Hoehn KL, Santos WL. "Small Molecule Mitochondrial Uncouplers and Their Therapeutic Potential". *Journal of medicinal chemistry*, 2018, 61 (11): 4641-4655, @2018 [Линк](#)
189. Gao W, Shi P, Chen X, Zhang L, Liu J, Fan X, Luo X. "Clathrin-mediated integrin  $\alpha\text{IIb}\beta\text{3}$  trafficking controls platelet spreading". *Platelets*. 29(6):610-21., @2018 [Линк](#)
190. Martinière A, Gibrat R, Sentenac H, Dumont X, Gaillard I, Paris N. "Uncovering pH at both sides of the root plasma membrane interface using noninvasive imaging". *Proceedings of the National Academy of Sciences of the United States of America*. 115 (25): 6488-6493., @2018 [Линк](#)
191. Okada Y, Ueda E, Kondo Y, Ishitsuka Y, Irie T, Higashi T, Motoyama K, Arima H, Matuso M, Higaki K, Ohno K. "Role of 6-O- $\alpha$ -maltosyl- $\beta$ -cyclodextrin in lysosomal cholesterol deprivation in Npc1-deficient Chinese hamster ovary cells". *Carbohydrate research*. 455, 54-61, @2018 [Линк](#)
192. Reynolds GD, Wang C, Pan J, Bednarek SY. Inroads into Internalization: Five Years of Endocytic Exploration. *Plant Physiology*, 176(1): pp. 208-218, @2018 [Линк](#)
193. Tejos R, Osorio-Navarro C, Norambuena L. "The Use of Drugs in the Study of Vacuole Morphology and Trafficking to the Vacuole in Arabidopsis thaliana". In: *Plant Vacuolar Trafficking*, pp. 143-154. Humana Press, New York, NY., @2018 [Линк](#)
194. De Vriese K, Himschoot E, Dünser K, Nguyen L, Drozdzecki A, Costa A, Nowack MK, Kleine-Vehn J, Audenaert D, Beeckman T, Vanneste S. "Identification of novel inhibitors of auxin-induced Ca<sup>2+</sup> signaling via a plant-based chemical screen". *Plant Physiology*. 2019; 180(1):480-96., @2019 [Линк](#)
195. Ekanayake G, LaMontagne ED, Heese A. "Never Walk Alone: Clathrin-Coated Vesicle (CCV) Components in Plant Immunity". *Annual Review of Phytopathology*. 2019; 57:387-409., @2019 [Линк](#)
196. Hu M, Schulze KE, Ghildyal R, Henstridge DC, Kolanowski JL, New EJ, Hong Y, Hsu AC, Hansbro PM, Wark PA, Bogoyevitch MA. "Respiratory syncytial virus co-opts host mitochondrial function to favour infectious virus production". *Elife*. 2019; 8., @2019 [Линк](#)
197. Huang L, Li X, Zhang C. "Progress in using chemical biology as a tool to uncover novel regulators of plant endomembrane trafficking". *Current Opinion in Plant Biology*. 2019; 52:106-13., @2019 [Линк](#)
198. Mazzon M, Marsh M. "Targeting viral entry as a strategy for broad-spectrum antivirals". *F1000Research*. 2019; 8., @2019 [Линк](#)
199. Mazzon M, Ortega-Prieto AM, Imrie D, Luft C, Hess L, Czieso S, Grove J, Skelton JK, Farleigh L, Bugert JJ, Wright E. "Identification of Broad-Spectrum Antiviral Compounds by Targeting Viral Entry". *Viruses*. 2019; 11(2):176., @2019 [Линк](#)
200. Ohbayashi I, Huang S, Fukaki H, Song X, Sun S, Morita MT, Tasaka M, Millar AH, Furutani M. "Mitochondrial pyruvate dehydrogenase contributes to auxin-regulated organ development". *Plant Physiology*. 2019; 180(2):896-909., @2019 [Линк](#)
201. Song JH, Kwak SH, Nam KH, Schiefelbein J, Lee MM. "QUIRKY regulates root epidermal cell patterning through stabilizing SCRAMBLED to control CAPRICE movement in Arabidopsis". *Nature Communications*. 2019; 10(1):1744., @2019 [Линк](#)
202. Słupianek A, Kasprowicz-Maluśki A, Myśkow E, Turzańska M, Sokołowska K. "Endocytosis acts as transport pathway in wood". *New Phytologist*. 2019; 222(4): 1846-61., @2019 [Линк](#)

203. Todkar K, Chikhi L, Germain M. "Mitochondrial interaction with the endosomal compartment in endocytosis and mitochondrial transfer". *Mitochondrion*. 2019 May 14., @2019 [Линк](#)
204. Zhang C, Hicks GR. "Braking plant endocytosis". *Nature Chemical Biology*. 2019; 15(6):553., @2019 [Линк](#)
205. Belda-Palazón B, Rodríguez PL. "Degradation of Abscisic Acid Receptors Through the Endosomal Pathway". In: *Plant Endosomes 2020* (pp. 35-48). Humana, New York, NY., @2020 [Линк](#)
206. De Caroli M, Manno E, Perrotta C, De Lorenzo G, Di Sansebastiano GP, Piro G. "CesA6 and PGIP2 Endocytosis Involves Different Subpopulations of TGN-Related Endosomes". *Frontiers in Plant Science*, 2020 Mar 27; 11:350., @2020 [Линк](#)
207. Deus CM, Yambire KF, Oliveira PJ, Raimundo N. "Mitochondria-lysosome crosstalk: from physiology to neurodegeneration". *Trends in Molecular Medicine*, 2020 Jan 1; 26(1):71-88., @2020 [Линк](#)
208. Foissner I, Hoefftberger M, Hoepflinger MC, Sommer A, Bulychev AA. "Brefeldin A inhibits clathrin-dependent endocytosis and ion transport in *Chara* internodal cells". *Biology of the Cell*, 2020 Jul 10., @2020 [Линк](#)
209. Ge FR, Chai S, Li S, Zhang Y. "Targeting and signaling of Rho of plants guanosine triphosphatases require synergistic interaction between guanine nucleotide inhibitor and vesicular trafficking". *Journal of Integrative Plant Biology*, 2020 Mar 21., @2020 [Линк](#)
210. Ju Y, Guo H, Edman M, Hamm-Alvarez SF. "Application of advances in endocytosis and membrane trafficking to drug delivery". *Advanced Drug Delivery Reviews*, 2020 Aug 3., @2020 [Линк](#)
211. Qi X, Yoshinari A, Bai P, Maes M, Zeng SM, Torii KU. "The manifold actions of signaling peptides on subcellular dynamics of a receptor specify stomatal cell fate". *Elife*, 2020 Aug 14; 9:e58097., @2020 [Линк](#)
212. Rajagopal D, Mathew MK. "Role of Arabidopsis RAB5 GEF vps9a in maintaining potassium levels under sodium chloride stress". *Plant Direct*, 2020 Oct; 4(10):e00273., @2020 [Линк](#)
213. Schwihla M, Korbei B. "The Beginning of the End: Initial Steps in the Degradation of Plasma Membrane Proteins". *Frontiers in Plant Science*, 2020 May 21; 11:680., @2020 [Линк](#)
214. Sun Y, Liang W, Cheng H, Wang H, Lv D, Wang W, Liang M, Miao C. "NADPH Oxidase-derived ROS promote mitochondrial alkalization under salt stress in Arabidopsis root cells". *Plant Signaling & Behavior*. 2020 Dec 14:1856546., @2020 [Линк](#)
215. Tran TM, Ma Z, Triebel A, Nath S, Cheng Y, Gong BQ, Han X, Wang J, Li JF, Wenk MR, Torta F. "The bacterial quorum sensing signal DSF hijacks Arabidopsis thaliana sterol biosynthesis to suppress plant innate immunity". *Life Science Alliance*, 2020, 3(10): e202000720., @2020 [Линк](#)
216. Zhu D, Zhang M, Gao C, Shen J. "Protein trafficking in plant cells: Tools and markers". *Science China Life Sciences*, 2020 Mar; 63(3):343-63., @2020 [Линк](#)
217. Zou C, Miffilin L, Hu Z, Zhang T, Shan B, Wang H, Xing X, Zhu H, Adiconis X, Levin JZ, Li F, Liu CF, Liu JS, Yuan J. "Reduction of mNAT1/hNAT2 Contributes to Cerebral Endothelial Necroptosis and Aβ Accumulation in Alzheimer's Disease". *Cell Reports*. 2020, 33(10): 108447., @2020 [Линк](#)
218. Ivanov R, Vert G. "Endocytosis in plants: Peculiarities and roles in the regulated trafficking of plant metal transporters". *Biology of the Cell*, 2021, 113(1):1-13., @2021 [Линк](#)
13. Betti C, Vanhoutte I, Coutuer S, De Rycke RM, **Mishev K**, Vuylsteke M, Aesaert S, Rombaut D, Gallardo R, De Smet F, Xu J, Van Lijsebettens M, Van Breusegem F, Inzé D, Rousseau F, Schymkowitz J, Russinova E. Sequence-specific protein aggregation generates defined protein knockdowns in plants. *Plant Physiology*, 171, American Society of Plant Biologists (ASPB), 2016, ISSN:1532-2548, Web of Science, DOI:10.1104/pp.16.00335, 773-787. ISI IF:6.456

Цитира се е:

219. Siddiqui MF, Bano B. "A biophysical insight into the formation of aggregates upon trifluoroethanol induced structural and conformational changes in garlic cystatin". *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 204: 7-17., @2018 [Линк](#)
220. Swarupini DS, Bhuyan AK. "Amyloid fibrillation of an intrinsically disordered plant phloem protein AtPP16-1 under acidic condition". *Biophysical chemistry*. 237:1-8., @2018 [Линк](#)

14. **Dimitrova AD**, Georgiev O, **Mishev K**, **Tzvetkov S**, Ananiev ED, Karagyozev L. Mapping of unmethylated sites in rDNA repeats in barley NOR deletion line. *Journal of Plant Physiology*, 205, Elsevier, 2016, ISSN:0176-1617, Web of Science, DOI:10.1016/j.jplph.2016.07.019, 97-104. ISI IF:3.121

Цитира се в:

221. A. Movahedi, J. Zhang, W. Sun, K. Mohammadi, A.A.Z. Yaghutia, H. Weia, X. Wua, T. Yinb, Q. Zhugea. Functional analyses of PtRDM1 gene overexpression in poplars and evaluation of its effect on DNA methylation and response to salt stress. *Plant Physiology and Biochemistry*, 2018, Vol. 127, 64–73., @2018 [Линк](#)

---

## 2018

---

15. **Mishev K**, Lu Q, Denoo B, Peurois F, Dejonghe W, Hullaert J, De Rycke R, Boeren S, Bretou M, De Munck S, Sharma I, Goodman K, Kalinowska K, Storme V, Nguyen LSL, Drozdzecki A, Martins S, Nerinckx W, Audenaert D, Vert G, Madder A, Otegui MS, Isono E, Savvides SN, Annaert W, De Vries S, Cherfils J, Winne J, Russinova E. Nonselective chemical inhibition of Sec7 domain-containing ARF GTPase exchange factors. *The Plant Cell*, 30, 10, American Society of Plant Biologists (ASPB), 2018, ISSN:1531-298X, Web of Science, DOI:10.1105/tpc.18.00145, 2573-2593. ISI IF:8.631

Цитира се в:

222. Depaepe T, Van Der Straeten D. "Tools of the Ethylene Trade: A Chemical Kit to Influence Ethylene Responses in Plants and Its Use in Agriculture". *Small Methods*. 2019 Aug 28., @2019 [Линк](#)
223. Huang L, Li X, Zhang C. "Progress in using chemical biology as a tool to uncover novel regulators of plant endomembrane trafficking". *Current Opinion in Plant Biology*. 2019; 52:106-13., @2019 [Линк](#)
224. Kerchev P, van der Meer T, Sujeeth N, Verlee A, Stevens CV, Van Breusegem F, Gechev T. "Molecular priming as an approach to induce tolerance against abiotic and oxidative stresses in crop plants". *Biotechnology Advances*. 2019: 107503., @2019 [Линк](#)
225. Huang L, Zhang C. "Perturbation and imaging of exocytosis in plant cells". *Methods in Cell Biology*, 2020; 160:3-20., @2020 [Линк](#)
226. Semerádova H, Montesinos JC, Benkova E. "All roads lead to auxin: post-translational regulation of auxin transport by multiple hormonal pathways". *Plant Communications*, 2020 Apr 22:100048., @2020 [Линк](#)
16. Kania U, Nodzynski T, Lu Q, Hicks GR, Nerinckx W, **Mishev K**, Peurois F, Cherfils J, De Rycke R, Groner P, Robert S, Russinova E, Friml J. The inhibitor Endosidin 4 targets SEC7 domain-type ARF GTPase exchange factors and interferes with subcellular trafficking in eukaryotes. *The Plant Cell*, 30, 10, American Society of Plant Biologists (ASPB), 2018, ISSN:1531-298X, Web of Science, DOI:10.1105/tpc.18.00127, 2553-2572. ISI IF:8.631

Цитира се в:

227. Huang L, Li X, Zhang C. "Progress in using chemical biology as a tool to uncover novel regulators of plant endomembrane trafficking". *Current Opinion in Plant Biology*. 2019; 52:106-13., @2019 [Линк](#)
228. Luzarowski M, Skirydz A. "Emerging strategies for the identification of protein–metabolite interactions". *Journal of Experimental Botany*. 2019; 70(18):4605-18., @2019 [Линк](#)
229. Semerádova H, Montesinos JC, Benkova E. "All roads lead to auxin: post-translational regulation of auxin transport by multiple hormonal pathways". *Plant Communications*, 2020 Apr 22:100048., @2020 [Линк](#)
230. Zhao Z, Yang X, Lü S, Fan J, Opiyo S, Yang P, Mangold J, Mackey D, Xia Y. "Deciphering the Novel Role of AtMIN7 in Cuticle Formation and Defense against the Bacterial Pathogen Infection". *International journal of molecular sciences*, 2020 Jan; 21(15):5547., @2020 [Линк](#)
17. Zhou J, Liu D, Wang P, Ma X, Lin W, Chen S, **Mishev K**, Lu D, Kumar R, Vanhoutte I, Meng X, He P, Russinova E, Shan L. Regulation of Arabidopsis brassinosteroid receptor BRI1 endocytosis and degradation by plant U-box PUB12/PUB13-mediated ubiquitination. *Proceedings of the National Academy of Sciences of the United States of America*, 115, 8, National Academy of Sciences of USA, 2018, ISSN:1091-6490, Web of Science, DOI:https://doi.org/10.1073/pnas.1712251115, E1906-E1915. ISI IF:9.58



Цитира се в:

231. Dai Vu L, Gevaert K, De Smet I. "Protein Language: Post-Translational Modifications Talking to Each Other". *Trends in plant science*, Vol. 23(12), p. 1068-1080., @2018 [Линк](#)
232. Gruszka D. "Crosstalk of the Brassinosteroid Signalosome with Phytohormonal and Stress Signaling Components Maintains a Balance between the Processes of Growth and Stress Tolerance". *International journal of molecular sciences*. 19(9):2675., @2018 [Линк](#)
233. Luo X, Liu J. Insights into receptor-like kinases-activated downstream events in plants. *Science China Life Sciences*, 61: 1586–1588., @2018 [Линк](#)
234. Lv M, Li M, Chen W, Wang Y, Sun C, Yin H, He K, Li J. "Thermal-enhanced bri1-301 instability reveals a plasma membrane protein quality control system in plants". *Frontiers in Plant Science*. 9: 1620., @2018 [Линк](#)
235. Qi X, Pleskot R, Irani NG, Van Damme D. "Meeting report–Cellular gateways: expanding the role of endocytosis in plant development." *J Cell Sci*. 131(17):jcs222604., @2018 [Линк](#)
236. Raboanatahiry N, Chao H, Dalin H, Pu S, Yan W, Yu L, Wang B, Li M. "QTL Alignment for Seed Yield and Yield Related Traits in Brassica napus". *Frontiers in plant science*. 9: 1127., @2018 [Линк](#)
237. Sharma C, Saripalli G, Kumar S, Gautam T, Kumar A, Rani S, Jain N, Prasad P, Raghuvanshi S, Jain M, Sharma JB. "A study of transcriptome in leaf rust infected bread wheat involving seedling resistance gene Lr28". *Functional Plant Biology*. 45(10):1046-64., @2018 [Линк](#)
238. Turek I, Tischer N, Lassig R, Trujillo M. "Multi-tiered pairing selectivity between E2 ubiquitin-conjugating enzymes and E3 ligases". *Journal of Biological Chemistry*. 293(42):16324-36., @2018 [Линк](#)
239. Chen Q, Chen QJ, Sun GQ, Zheng K, Yao ZP, Han YH, Wang LP, Duan YJ, Yu DQ, Qu YY. "Genome-wide identification of cyclophilin gene family in cotton and expression analysis of the fibre development in *Gossypium barbadense*". *International Journal of Molecular Sciences*. 2019; 20(2):349., @2019 [Линк](#)
240. Copeland C, Li X. "Regulation of Plant Immunity by the Proteasome". *International Review of Cell and Molecular Biology*. 2019; 343:37-63., @2019 [Линк](#)
241. Gao J, Chaudhary A, Vaddepalli P, Nagel MK, Isono E, Schneitz K. "The Arabidopsis receptor kinase STRUBBELIG undergoes clathrin-dependent endocytosis". *Journal of Experimental Botany*. 2019 Apr 25., @2019 [Линк](#)
242. Hussain S, Gomes MM, Yano K, Nambara E. "Interactions between abscisic acid and other hormones". *Abscisic Acid in Plants*. 2019; 24:255., @2019 [Линк](#)
243. Lin L, Zhang C, Chen Y, Wang Y, Wang D, Liu X, Wang M, Mao J, Zhang J, Xing W, Liu L. "PAWH1 and PAWH2 are plant-specific components of an Arabidopsis endoplasmic reticulum-associated degradation complex". *Nature Communications*. 2019; 10(1):1-3., @2019 [Линк](#)
244. McKenna JF, Rolfe DJ, Webb SE, Tolmie AF, Botchway SW, Martin-Fernandez ML, Hawes C, Runions J. "The cell wall regulates dynamics and size of plasma-membrane nanodomains in Arabidopsis". *Proc Natl Acad Sci U S A*. 2019; 116(26):12857-62., @2019 [Линк](#)
245. Minamino N, Ueda T. "RAB GTPases and their effectors in plant endosomal transport". *Current Opinion in Plant Biology*. 2019; 52:61-8., @2019 [Линк](#)
246. Rodriguez-Furlan C, Minina EA, Hicks GR. "Remove, Recycle, Degrade: Regulating Plasma Membrane Protein Accumulation". *The Plant Cell*. 2019; 31(12):2833-54., @2019 [Линк](#)
247. Uhrig RG, Schläpfer P, Roschitzki B, Hirsch-Hoffmann M, Gruissem W. "Diurnal changes in concerted plant protein phosphorylation and acetylation in Arabidopsis organs and seedlings". *The Plant Journal*. 2019 Mar 28., @2019 [Линк](#)
248. Wang J, Liu S, Liu H, Chen K, Zhang P. "PnSAG1, an E3 ubiquitin ligase of the Antarctic moss *Pohlia nutans*, enhanced sensitivity to salt stress and ABA". *Plant Physiology and Biochemistry*. 2019 Jun 5., @2019 [Линк](#)
249. Zhang L, Liu Y, Zhu XF, Jung JH, Sun Q, Li TY, Chen LJ, Duan YX, Xuan YH. "SYP22 and VAMP727 regulate BRI1 plasma membrane targeting to control plant growth in Arabidopsis". *New Phytologist*. 2019 Feb 25., @2019 [Линк](#)
250. Aguilar-Hernandez V, Brito-Argaez L, Galaz-Avalos RM, Loyola-Vargas VM. "Post-translational modifications drive plant cell differentiation". *Plant Cell, Tissue and Organ Culture*, 2020 Aug 9:1-2., @2020 [Линк](#)

251. Feke AM, Hong J, Liu W, Gendron JM. "A Decoy Library Uncovers U-box E3 Ubiquitin Ligases that Regulate Flowering Time in Arabidopsis". *Genetics*, 2020 May 20., @2020 [Линк](#)
252. Galindo-Trigo S, Blümke P, Simon R, Butenko MA. "Emerging mechanisms to fine-tune receptor kinase signaling specificity". *Current Opinion in Plant Biology*, 2020 Oct 1; 57:41-51., @2020 [Линк](#)
253. Johnson A, Gnyliukh N, Kaufmann WA, Narasimhan M, Vert G, Bednarek SY, Friml J. "Experimental toolbox for quantitative evaluation of clathrin-mediated endocytosis in the plant model Arabidopsis". *Journal of Cell Science*, 2020 Aug 1; 133(15)., @2020 [Линк](#)
254. Kumar V, Donev EN, Barbut FR, Kushwah S, Mannapperuma C, Urbancsok J, Mellerowicz EJ. "Genome-wide identification of Populus malectin/malectin-like domain-containing proteins and expression analyses reveal novel candidates for signaling and regulation of wood development". *Frontiers in Plant Science*, 2020; 11: 588846., @2020 [Линк](#)
255. Lan Y, Wu L, Wu M, Liu H, Gao Y, Zhang K, Xiang Y. "Transcriptome analysis reveals key genes regulating signaling and metabolic pathways during the growth of moso bamboo (*Phyllostachys edulis*) shoots". *Physiologia Plantarum*. 2020 Dec 5, DOI: 10.1111/ppl.13296., @2020 [Линк](#)
256. Lee D, Lal NK, Lin ZJ, Ma S, Liu J, Castro B, Toruño T, Dinesh-Kumar SP, Coaker G. "Regulation of reactive oxygen species during plant immunity through phosphorylation and ubiquitination of RBOHD". *Nature Communications*, 2020 Apr 15; 11(1):1-6., @2020 [Линк](#)
257. Li Q, Serio RJ, Schofield A, Liu H, Rasmussen SR, Hofius D, Stone SL. "Arabidopsis RING-type E3 ubiquitin ligase XBAT35. 2 promotes proteasome-dependent degradation of ACD11 to attenuate abiotic stress tolerance". *The Plant Journal*, 2020 Oct 20., @2020 [Линк](#)
258. Linden KJ, Callis J. "The ubiquitin system affects agronomic plant traits". *Journal of Biological Chemistry*, 2020 Oct 2; 295(40):13940-55., @2020 [Линк](#)
259. Lv M, Li J. "Molecular Mechanisms of Brassinosteroid-Mediated Responses to Changing Environments in Arabidopsis". *International Journal of Molecular Sciences*. 2020 Jan; 21(8):2737., @2020 [Линк](#)
260. Mao J, Li J. "Regulation of Three Key Kinases of Brassinosteroid Signaling Pathway". *International Journal of Molecular Sciences*, 2020 Jan; 21(12):4340., @2020 [Линк](#)
261. Qi X, Yoshinari A, Bai P, Maes M, Zeng SM, Torii KU. "The manifold actions of signaling peptides on subcellular dynamics of a receptor specify stomatal cell fate". *Elife*, 2020 Aug 14; 9:e58097., @2020 [Линк](#)
262. Sun F, Yu H, Qu J, Cao Y, Ding L, Feng W, Khalid MH, Li W, Fu F. "Maize ZmBES1/BZR1-5 Decreases ABA Sensitivity and Confers Tolerance to Osmotic Stress in Transgenic Arabidopsis". *International Journal of Molecular Sciences*, 2020 Jan; 21(3):996., @2020 [Линк](#)
263. Wang M, Li X, Luo S, Fan B, Zhu C, Chen Z. "Coordination and Crosstalk between Autophagosome and Multivesicular Body Pathways in Plant Stress Responses". *Cells*, 2020 Jan; 9(1):119., @2020 [Линк](#)
264. Wang Q, Yu F, Xie Q. "Balancing growth and adaptation to stress: Crosstalk between brassinosteroid and abscisic acid signaling". *Plant, Cell & Environment*, 2020 Oct; 43(10):2325-35., @2020 [Линк](#)
265. Wu Z, Tong M, Tian L, Zhu C, Liu X, Zhang Y, Li X. "Plant E3 ligases SNIPER 1 and SNIPER 2 broadly regulate the homeostasis of sensor NLR immune receptors". *The EMBO Journal*, 2020 Aug 3; 39(15):e104915., @2020 [Линк](#)
266. Chen X, Wang T, Rehman AU, Wang Y, Qi J, Li Z, Song C, Wang B, Yang S, Gong Z. "Arabidopsis U-box E3 ubiquitin ligase PUB11 negatively regulates drought tolerance by degrading the receptor-like protein kinases LRR1 and KIN7". *Journal of Integrative Plant Biology*, 2021, 63(3): 494-509., @2021 [Линк](#)
267. Miao R, Yuan W, Wang Y, Garcia-Maquilon I, Dang X, Li Y, Zhang J, Zhu Y, Rodriguez PL, Xu W. "Low ABA concentration promotes root growth and hydrotropism through relief of ABA INSENSITIVE 1-mediated inhibition of plasma membrane H<sup>+</sup>-ATPase 2". *Science Advances*, 2021, 7(12): eabd4113., @2021 [Линк](#)
268. Nazir F, Fariduddin Q, Hussain A, Khan TA. "Brassinosteroid and hydrogen peroxide improve photosynthetic machinery, stomatal movement, root morphology and cell viability and reduce Cu-triggered oxidative burst in tomato". *Ecotoxicology and Environmental Safety*, 2021, 207: 111081., @2021 [Линк](#)
269. Turek I, Irving H. "Moonlighting proteins shine new light on molecular signaling niches". *International Journal of Molecular Sciences*, 2021, 22(3): 1367., @2021 [Линк](#)

18. Kirilova I, Hristeva T, Bozhinova R, **Mishev K, Vassileva V, Dimitrova A**, Denev I. Plant parasitism from the prospective of the system host plants-parasitic broomrapes-soil microbiome. International congress on oil and protein crops, 2018, 108

Цитира се в:

270. Lyakh VA, Kostyuchenko NI, Shevchenko IA. Broomrape (*Orobanche cumana* Wallr.) can influence the microbial cenosis in sunflower rhizosphere. *Helia* 42, 71, 145-159, @2019 [Линк](#)

---

## 2019

---

19. Dejonghe W, Sharma I, Denoo B, De Munck S, Lu Q, **Mishev K**, Bulut H, Mylle E, De Rycke R, Vasileva M, Savatin DV, Nerinckx W, Staes A, Drozdzecki A, Audenaert D, Yperman K, Madder A, Friml J, Van Damme D, Gevaert K, Haucke V, Savvides SN, Winne J, Russinova E. Disruption of endocytosis through chemical inhibition of clathrin heavy chain function. *Nature Chemical Biology*, 15, 6, Springer Nature Publishing AG, 2019, ISSN:1552-4469, Web of Science, DOI:10.1038/s41589-019-0262-1, 641-649. JCR-IF (Web of Science):12.587

Цитира се в:

271. Huang L, Li X, Zhang C. "Progress in using chemical biology as a tool to uncover novel regulators of plant endomembrane trafficking". *Current Opinion in Plant Biology*. 2019; 52:106-13., @2019 [Линк](#)
272. Belda-Palazón B, Rodriguez PL. "Degradation of Abscisic Acid Receptors Through the Endosomal Pathway". In: *Plant Endosomes*, 2020, pp. 35-48. Humana, New York, NY., @2020 [Линк](#)
273. Ha J, Park H, Park J, Park SB. "Recent advances in identifying protein targets in drug discovery". *Cell Chemical Biology*. 2020 Dec 22, DOI: 10.1016/j.chembiol.2020.12.001., @2020 [Линк](#)
274. Iwatate RJ, Yoshinari A, Yagi N, Grzybowski M, Ogasawara H, Kamiya M, Komatsu T, Taki M, Yamaguchi S, Frommer WB, Nakamura M. "Covalent self-labeling of tagged proteins with chemical fluorescent dyes in BY-2 cells and Arabidopsis seedlings". *The Plant Cell*, 2020; 32(10):3081-94., @2020 [Линк](#)
275. Ju Y, Guo H, Edman M, Hamm-Alvarez SF. "Application of advances in endocytosis and membrane trafficking to drug delivery". *Advanced Drug Delivery Reviews*, 2020 Aug 3., @2020 [Линк](#)
276. Qi X, Yoshinari A, Bai P, Maes M, Zeng SM, Torii KU. "The manifold actions of signaling peptides on subcellular dynamics of a receptor specify stomatal cell fate". *Elife*, 2020; 9:e58097., @2020 [Линк](#)
277. Schwihla M, Korbei B. "The Beginning of the End: Initial Steps in the Degradation of Plasma Membrane Proteins". *Frontiers in Plant Science*, 2020; 11:680., @2020 [Линк](#)
278. Semerádova H, Montesinos JC, Benkova E. "All roads lead to auxin: post-translational regulation of auxin transport by multiple hormonal pathways". *Plant Communications*, 2020 Apr 22:100048., @2020 [Линк](#)
279. Tobys D, Kowalski LM, Cziudaj E, Müller S, Zentis P, Pach E, Zigrino P, Blaeske T, Höning S. "Inhibition of clathrin-mediated endocytosis by knockdown of AP-2 leads to alterations in the plasma membrane proteome". *Traffic*. 2020 Nov 22, DOI: 10.1111/tra.12770., @2020 [Линк](#)
280. Tran TM, Ma Z, Triebel A, Nath S, Cheng Y, Gong BQ, Han X, Wang J, Li JF, Wenk MR, Torta F. "The bacterial quorum sensing signal DSF hijacks Arabidopsis thaliana sterol biosynthesis to suppress plant innate immunity". *Life Science Alliance*, 2020, 3(10): e202000720, @2020 [Линк](#)
281. de Almeida MS, Susnik E, Drasler B, Taladriz-Blanco P, Petri-Fink A, Rothen-Rutishauser B. "Understanding nanoparticle endocytosis to improve targeting strategies in nanomedicine". *Chemical Society Reviews*. 2021, DOI: 10.1039/D0CS01127D, @2021 [Линк](#)
282. Li X, Zhang C. "Using Differential Scanning Fluorimetry (DSF) to Detect Ligand Binding with Purified Protein". In: *Plant Chemical Genomics*. 2021, pp. 183-186, Humana, New York, NY., @2021 [Линк](#)
283. Rodriguez-Furlan C, Hicks GR. "Label-Free Target Identification and Confirmation Using Thermal Stability Shift Assays". In: *Plant Chemical Genomics*. 2021, pp. 163-173, Humana, New York, NY., @2021 [Линк](#)
284. Shirakawa M, Morisaki Y, Gan ES, Sato A, Ito T. "Identification of a devernalization inducer by chemical screening approaches in Arabidopsis thaliana". *Frontiers in plant science*, 2021, 12: 634068., @2021 [Линк](#)

285. Tobys D, Kowalski LM, Cziudaj E, Müller S, Zentis P, Pach E, Zigrino P, Blaeske T, Höning S. "Inhibition of clathrin-mediated endocytosis by knockdown of AP-2 leads to alterations in the plasma membrane proteome". Traffic, 2021, 22(1-2): 6-22., @2021 [Линк](#)

16.04.2021 г.  
гр. София

Подпис:  
(Кирил Мишев)