

**ЦИТАТИ**  
(без автоцитати и полуцитати)  
Гл. ас. д-р Мария Иванова Петкова

**СПРАВКА ВСИЧКИ ЦИТИРАНИЯ**

Вид на цитиращото издание: всички издания

Година: 2005-2025

Брой цитирани публикации: 39	Брой цитиращи източници: 425
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**СПИСЪК НА ЦИТИРАНИЯ В SCOPUS ИЛИ WEB OF SCIENCE**

Вид на цитиращото издание: Публикации, индексирани в Scopus или Web of Science

Година: 2005-2025

Брой цитирани публикации: 37	Брой цитиращи източници: 252
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**СПИСЪК С ЦИТИРАНИЯ ЗА УЧАСТИЕ В КОНКУРСА**

(извадка от последните 5 години)

Вид на цитиращото издание: Публикации, индексирани в Scopus или Web of Science

Година: 2021-2025

Брой цитирани публикации: 35	Брой цитиращи източници: 174
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2006

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1. **Petrova, M**, Zagorska, N, Tasheva, K, Evstatieva, L. *In vitro propagation of Gentiana lutea*. L. Genetics and Breeding, 35, 1-2, 63-68, 2006

Цитира се в:

1. Aras Aşçı, Ö., Demirci, T., Albayrak, İ., Deveci, H., & Göktürk Baydar, N. Optimization of inoculum density to support root growth and secondary metabolite accumulation in root cultures of endangered *Gentiana* species: *Gentiana lutea* and

*Gentiana boissieri*. In Vitro Cellular & Developmental Biology-Plant, 58, 1090–1098, 2022., @2022 [Линк](#)

2. Demirci, T., Albayrak, I., Deveci, H., Asci, Ö. A., Baydar, N. G. "Development of effective micropropagation protocols for endangered *Gentiana lutea* and *Gentiana boissieri* (endemic to Türkiye)." Israel Journal of Plant Sciences. 71(3-4), 127-136, 2024 <https://doi.org/10.1163/22238980-bja10108>, @2024 [Линк](#)
2. **Petrova M.**, Stoilova T, Zagorska N. Isoenzyme and protein patterns of *in vitro* micropropagated plantlets of *Gentiana lutea* L. after application of various growth regulators. Biotechnology & Biotechnological Equipment, 20(1), 15-19, 2006

Цитира се в:

3. Aday Kaya , A. G., Albayrak, İ., Demirci, T., Deveci, H., Baydar, N. G. "MeJA Changes Root Growth, Iridoid, Xanthone, and Secoiridoid Production, as well as Gene Expression Levels in Root Cultures of Endangered *Gentiana lutea* and *Gentiana boissieri*." Journal of Plant Growth Regulation, 44, 295–315, 2025, @2025 [Линк](#)

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## 2008

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3. **Petrova M.**, Zayova E., Vlahova M. Induction of callus cultures in *Arnica montana*. Genetics and Breeding, 37, 3-4, 2008, 37-34

Цитира се в:

4. Nieto-Trujillo, A., Cruz-Sosa, F., Luria-Pérez, R.; Gutiérrez-Rebolledo, G.A., Román-Guerrero, A., Burrola-Aguilar, C., Zepeda-Gómez, C., Estrada-Zúñiga, M.E. *Arnica montana* Cell Culture Establishment, and Assessment of Its Cytotoxic, Antibacterial,  $\alpha$ -Amylase Inhibitor, and Antioxidant In Vitro Bioactivities. Plants 2021, 10, 2300. <https://doi.org/10.3390/plants10112300>, @2021 [Линк](#)

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## 2011

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4. **Petrova, M.**, Zayova, E., Yankova, E., Baldzhiev, G..Plant regeneration from callus culture of *Arnica montana*. Romanian Biotechnological Letters, 16(1), 92-97, 2011

Цитира се във:

5. Nieto-Trujillo, A., Cruz-Sosa, F., Luria-Pérez, R., Gutiérrez-Rebolledo, G.A., Román-Guerrero, A., Burrola-Aguilar, C., Zepeda-Gómez, C., Estrada-Zúñiga, M.E. *Arnica montana* Cell Culture Establishment, and Assessment of Its Cytotoxic, Antibacterial,  $\alpha$ -Amylase Inhibitor, and Antioxidant In Vitro Bioactivities. *Plants* 2021, 10, 2300. <https://doi.org/10.3390/plants10112300>, @2021 [Линк](#)
6. Saritha, K., Sandhya, D., Thirupathi, K., Mohammed, M. "Direct regeneration from leaf explants and genetic fidelity analysis of regenerates through ISSR markers in *Kedrostis foetidissima*: a medicinal climber. " *Vegetos*, 37(4), 1669-1676, 2024., @2024 [Линк](#)
5. **Petrova, M.**, Zayova, E., Vitkova, A. Effect of silver nitrate on *in vitro* root formation of *Gentiana lutea*. *Romanian Biotechnological Letters*, 16(6), 53-58, 2011.

Цитира се във:

7. Kara, Z., Sabır, A., Koç, F., Sabır, F. K., Avcı, A., Koplay, M., & Doğan, O. Silver Nanoparticles Synthesis by Grape Seeds (*Vitis vinifera* L.) Extract and Rooting Effect on Grape Cuttings. *Erwerbs-Obstbau* 63, 1–8 (2021). <https://doi.org/10.1007/s10341-021-00572-8>, @2021 [Линк](#)
8. Hoang, N. H., Le Thanh, T., Thepbandit, W., Treekoon, J., Saengchan, C., Sangpueak, R., Papathoti NK, Kamkaew A, Buensanteai, N. Efficacy of Chitosan Nanoparticle Loaded-Salicylic Acid and-Silver on Management of Cassava Leaf Spot Disease. *Polymers*, 14(4), 660, 2022., @2022 [Линк](#)
9. Isah, T., & Umar, S. Influence of silver nitrate and copper sulfate on somatic embryogenesis, shoot morphogenesis, multiplication, and associated physiological biochemical changes in *Gladiolus hybridus* L. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 149, 563–587, 2022., @2022 [Линк](#)
10. Srivastava, S., Nalla, V. K., Singh, V. P., & Prasad, R. Optimization of *in vitro* micropropagation and root establishment through combinatorial approaches for enhanced production of secondary metabolites in the endangered species *Decalepis aryalpathra* KMA 05 clones. *Brazilian Journal of Botany*, 45(3), 869-881, 2022., @2022 [Линк](#)

11. Feizi, S. Role of Nanomaterials in Plant Cell and Tissue Culture. In: Al-Khayri, J.M., Alnaddaf, L.M., Jain, S.M. (eds) Nanomaterial Interactions with Plant Cellular Mechanisms and Macromolecules and Agricultural Implications. Springer, Cham, 2023. [https://doi.org/10.1007/978-3-031-20878-2\\_14](https://doi.org/10.1007/978-3-031-20878-2_14), @2023 [Линк](#)
12. Demirci, T., Albayrak, I., Deveci, H., Asci, Ö. A., Baydar, N. G. "Development of effective micropropagation protocols for endangered *Gentiana lutea* and *Gentiana boissieri* (endemic to Türkiye)." Israel Journal of Plant Sciences, 71(3-4), 127-136, 2024, @2024 [Линк](#)
13. Kim, J. B., Kim, N. S., Lim, J., Kim, K., Lee, M., Uddin, M. R., Sathasivam, R., Park, C., & Park, S. U. Influences of carbon sources and plant growth regulators on *in vitro* rooting of *Lycium chinense*. Journal of Phytology, 17, 25–29, 2025, @2025 [Линк](#)

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## 2012

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6. Zayova, E, Stancheva, I, Geneva, M, **Petrova, M**, Vasilevska-Ivanova, R. Morphological evaluation and antioxidant activity of *in vitro*- and *in vivo*- derived *Echinacea purpurea* plants. Central European Journal of Biology, 7(4), 698-707, 2012.

Цитира се в:

14. Ahmadi F, Samadi A, Sepehr E, Rahimi A, Shabala S, 2022, Cell-type-specific H+-ATPase activity and antioxidant enzymes improve the *Echinacea purpurea* L. Moench tolerance to salinity stress at different NO<sub>3</sub>-/NH<sub>4</sub><sup>+</sup> ratios, Industrial Crops and Products, 186, 115199, ISSN 0926-6690, @2022 [Линк](#)
15. Tourky S., Shukry W., Hossain M., Siddiqui M., Pessarakli M., Elghareeb E. Cobalt enhanced the drought-stress tolerance of rice (*Oryza sativa* L.) by mitigating the oxidative damage and enhancing yield attributes, South African Journal of Botany, 159, 191-207, ISSN 0254-6299, @2023 [Линк](#)
16. Woch N, Laha S, Gudipalli P. Salicylic acid and jasmonic acid induced enhanced production of total phenolics, flavonoids, and antioxidant metabolism in callus cultures of *Givotia moluccana* (L.) Sreem. In Vitro Cellular & Developmental Biology-Plant, 59, 227–248 , 2023, @2023 [Линк](#)
17. Grzelak M, Pacholczak A, Nowakowska K. "The effect of several growth regulators and biostimulant on biochemical and physiological changes in acclimation of

micropagated *Echinacea purpurea* Moench. 'Raspberry Truffle". Plant Cell Tiss Organ Cult 159, 22, @2024 [Линк](#)

18. Wafa' Nur H, Ahmad Y, Nandariyah; Yuli W, "Morphological, agronomic characteristics, and flavonoid content of *Echinacea purpurea* at various gamma ray doses", Bulgarian Journal of Agricultural Science, 30(3), 451-547, @2024

7. **Petrova M**, Zayova E, Vassilevska-Ivanova R, Vlahova M. Biotechnological approaches for cultivation and enhancement of secondary metabolites in *Arnica montana* L. Acta Physiologiae Plantarum, 34, 1597-1606, 2012.

Цитира се във:

19. Nieto-Trujillo A, Cruz-Sosa F, Luria-Pérez R, Gutiérrez-Rebolledo GA, Román-Guerrero A, Burrola-Aguilar C, Zepeda-Gómez C, Estrada-Zúñiga ME. *Arnica montana* cell culture establishment, and assessment of its cytotoxic, antibacterial, α-amylase inhibitor, and antioxidant in vitro bioactivities. Plants 10, 11, 2300, @2021 [Линк](#)
20. Danna, C., Poggio, L., Smeriglio, A., Mariotti, M., & Cornara, L. Ethnomedicinal and Ethnobotanical Survey in the Aosta Valley Side of the Gran Paradiso National Park (Western Alps, Italy). Plants, 11(2), 170, 2022., @2022 [Линк](#)
21. Klimek-Szczykutowicz, M., Ekiert, H., Szopa, A. Biotechnological Studies on *Nasturtium officinale* (Watercress): an Endangered Species of Significant Relevance in Medicine, Cosmetic, and Food Industries. In Medicinal Plants: Biodiversity, Biotechnology and Conservation (pp. 595-631). Singapore: Springer Nature Singapore, 2023. @2023 [Линк](#)
22. Michalak, M., Błońska-Sikora, E. M., Paradowska, K., Zielińska, A. Pharmaceutical availability of hydrogels with extracts of *Arnica montana*, *Aesculus hippocastanum* and *Ruscus aculeatus* and their potential use as antioxidant polyphenol-rich material. Medical Studies/Studia Medyczne, 39(3), 223-229., @2023 [Линк](#)
23. Capria, L., Liepelt, S., Eimert, K., Leyer, I., Mosner, E. "Neutral genetic diversity follows a latitudinal gradient in the endangered plant *Arnica montana* L.: a range-wide study." Conservation Genetics, 25(1), 87-100, 2024, @2024 [Линк](#)

8. Zayova, E, Stancheva, I, Geneva, M, **Petrova, M**, Dimitrova, L. Antioxidant activity of in vitro propagated *Stevia rebaudiana* plants from different origins. Turkish Journal of Biology, 37(1), 106-113, 2013.

Цитира се е:

24. Dinev T., Tzanova M., Rusenova N., Grozева N., Gerdzhikova M., Beev G. Antimicrobial and antioxidant potential of methanolic extracts from different parts of *Stevia rebaudiana* Bertoni cultivated in Bulgaria. Sains Malaysiana 50(9): 2641-2651, @2021 [Линк](#)
25. Dinev, T., Tzanova, M., Velichkova, K., Dermendzhieva, D., & Beev, G. Antifungal and Antioxidant Potential of Methanolic Extracts from *Acorus calamus* L., *Chlorella vulgaris* Beijerinck, *Lemna minuta* Kunth and *Scenedesmus dimorphus* (Turpin) Kützing. Applied Sciences, 11(11), 4745, @2021 [Линк](#)
26. Mehmed, A., Enchev, S. & Bozhimirov, S. Assessment of initial material for stevia selection (*Stevia rebaudiana* B.). Bulg. J. Agric. Sci., 27 (3), 536-540, 2021, @2021 [Линк](#)
27. Fayed, A., Ramadan, H. O., Hassan, S. A., Hussein, M. A., & Roshdy, T. Thioredoxin1 Gene Modulates Bcl2/p53/NF-KB Signaling Pathways in Strawberry Extract/Paracetamol-treated Rat Model of Acute Liver Injury. Biomedical and Pharmacology Journal, 15(2), 1025-1037, 2022., @2022 [Линк](#)
28. Inceer H, Cuce M, Imamoglu KV, Ergin T, Ucler AO, 2022, In vitro propagation and cytogenetic stability of *Tripleurospermum insularum* (Asteraceae) – a critically endangered insular endemic species from Turkey, Plant Biosystems - An International Journal Dealing with all Aspects of Plant Biology, 156(5), 1213–1221, @2022 [Линк](#)
29. Rezvankhah M, Askari H, Tohidfar M, Rezadoost H, Economic micropropagation of *Stevia rebaudiana* Bertoni and evaluation of in vitro cultures in order to improve steviol glycosides, Scientia Horticulturae, 305, 111372, @2022 [Линк](#)
30. Bebek Markovinović A, Milošević S, Teslić N, Pavlić B, Putnik P, Brčić Karačonji I, Jurica K, Lasić D, Bursać Kovačević D. Development of a Pressurized Green Liquid

Extraction Procedure to Recover Antioxidant Bioactive Compounds from Strawberry Tree Fruit (*Arbutus unedo* L.). Plants. 12(10):2006, @2023 [Линк](#)

31. Özyigit Y, Uçar E, Erugur N, Ataş M, İnanir M, Bal H, Kahrizi D, Turgut K. Comparison of different drying methods for phytochemical quality of stevia (*Stevia rebaudiana* Bert.). Notulae Scientia Biologicae, 15(3), 11527-11527, @2023 [Линк](#)

32. Wijayanti E., Safitri A., Siswanto D., Fatchiyah F. Indonesian purple rice ferulic acid as a candidate for anti-aging through the inhibition of collagenase and tyrosinase activities. Indonesian Journal of Chemistry, 23(2), 475-488, @2023 [Линк](#)

9. Nikolova, M., Vitkova, A., Zayova, E., **Petrova, M.** Flavonoid Profiles, Polyphenolic content and antiradical properties of cultivated plants of *Arnica montana* L. AgroLife Scientific Journal, 2 (2), 20-24, 2013.

Цитира се въз:

33. Aiello, N., Fusani, P. and Dall'Acqua, S. Cultivation trial of *Arnica montana* L. wild populations compared with cv. 'Arbo", Italus Hortus, 28(2), pp. 70-78. doi: 10.26353/j.itahort/2021.2.7078, 2021, @2021 [Линк](#)

34. Darwish H., Al-Osaimi G.S., Al Kashgry N.A.T., Sonbol H., Alayafi A.A.M., Alabdallah N.M., Al-Humaid A., Al-Harbi N.A., Al-Qahtani S.M., Abbas Z.K., Darwish D.B.E, Ibrahim M.F.M., Noureldeen A. Evaluating the genotoxicity of salinity stress and secondary products gene manipulation in lime, *Citrus aurantifolia*, plants. Frontiers in Plant Science. 2023 Jul 12;14:1211595. doi:10.3389/fpls.2023.1211595., @2023 [Линк](#)

10. **Petrova, M.**, Zayova, E., Vlahova, M.. Induction of hairy roots in *Arnica montana* L. by *Agrobacterium rhizogenes*. Central European Journal of Biology, 8, 470-479, 2013.

Цитира се въз:

35. Cao, D. M., Tran, T. T., & Quach, P. N. Investigation of factors in improving Agrobacterium-mediated gene transfer in *Ruellia tuberosa* L. and evaluation of α-glucosidase inhibitory activity in established hairy roots. Notulae Botanicae Horti Agrobotanici Cluj-Napoca, 50(3), 12588, 2022, @2022 [Линк](#)

36. Zhao, S. Overexpression of geraniol-10-hydroxylase improves valtrate accumulation in *Valeriana jatamansi*. Plant Genetic Resources, 21(1), 45-49, 2023., [@2023](#) [Линк](#)
37. Barrera Núñez, M. G., Bueno, M., Molina-Montiel, M. Á., Reyes-Vaquero, L., Ibáñez, E., Del Villar-Martínez, A. A. Chemical Profile of Cell Cultures of *Kalanchoë gastonis-bonnieri* Transformed by *Agrobacterium rhizogenes*. Agronomy, 14(1), 189, 2024, [@2024](#) [Линк](#)
11. Nikolova, M., **Petrova, M.**, Zayova, E. Comparative study of in vitro, ex vitro and in vivo grown plants of *Arnica montana* – polyphenols and free radical scavenging activity. Acta botanica Croatica, 72, 1, 13-22, 2013.

Цитира се въз:

38. Flórez-Fernández, N., Ferreira-Anta, T., Torres, M. D., & Domínguez, H. Valorization of *Arnica montana* wastes after extraction of the ethanol tincture: application in polymer-based matrices. Polymers, 13(18), 3121., 2021, [@2021](#) [Линк](#)
39. Lei, T., Pan, Y., Zhang, B., Liu, R., & Pan, Y. Optimisation of ultrasonic-assisted extraction of natural dyes from pomegranate rind using response surface methodology and its characterisation. Coloration Technology, 137(3), 259-271, 2021, [@2021](#) [Линк](#)
40. Mangena P. Effect of water stress on growth responses of soybean plants pretreated with colchicine. Acta Hortic. 1327. ISHS 2021. DOI 10.17660/ActaHortic.2021.1327.15 Proc. IV International Symposium on Horticulture in Europe – SHE2021 Eds.: J.N. Wünsche and A. Milyaev, 2021., [@2021](#) [Линк](#)
41. Mangena, P. Effect of *Agrobacterium* co-cultivation stage on explant response for subsequent genetic transformation in Soybean (*Glycine max* (L.) Merr.). Plant Science Today, 8(4): 905-911., 2021, [@2021](#) [Линк](#)
42. Mangena, P. Germination, Morphological and Physiological Evaluation of Seedlings Pretreated with Colchicine in Soybean (*Glycine max* L). Walailak Journal of Science and Technology (WJST), 18(18), 9489-12, 2021., [@2021](#) [Линк](#)
43. Mantovska, D.I.; Zhiponova, M.K.; Georgiev, M.I.; Grozdanova, T.; Gerginova, D.; Alipieva, K.; Simova, S.; Popova, M.; Kapchina-Toteva, V.M.; Yordanova, Z.P. In

*Vitro* Multiplication and NMR Fingerprinting of Rare *Veronica caucasica* M. Bieb. Molecules, 26(19), 5888, 2021, [@2021](#) [Линк](#)

44. Nieto-Trujillo, A.; Cruz-Sosa, F.; Luria-Pérez, R.; Gutiérrez-Rebolledo, G.A.; Román-Guerrero, A.; Burrola-Aguilar, C.; Zepeda-Gómez, C.; Estrada-Zúñiga, M.E. *Arnica montana* Cell Culture Establishment, and Assessment of Its Cytotoxic, Antibacterial, α-Amylase Inhibitor, and Antioxidant *In Vitro* Bioactivities. Plants, 10(11), 2300, 2021, [@2021](#) [Линк](#)
45. Smith, A. G., Miles, V. N., Holmes, D. T., Chen, X., & Lei, W. Clinical Trials, Potential Mechanisms, and Adverse Effects of *Arnica* as an Adjunct Medication for Pain Management. Medicines, 8(10), 58., 2021., [@2021](#) [Линк](#)
46. Crisóstomo-Ayala, K. A., Sabater-Jara, A. B., Pérez Manríquez, C., Ferreres, F., Gil-Izquierdo, Á., Pedreño, M. Á., Hernández de la Torre, M., Sanchez-Olate, M., & Ríos Leal, D. G. Comparative Study of Metabolomic Profile and Antioxidant Content of Adult and *In Vitro* Leaves of *Aristotelia chilensis*. Plants, 11(1), 37, 2022 <https://doi.org/10.3390/plants11010037>, [@2022](#) [Линк](#)
47. Faraloni C., Giordano C., Arcidiaco L., Benelli C., Di Lonardo S., Anichini M., Stefani F., Petruccelli R. Effective Microorganisms and Olive Mill Wastewater Used as Biostimulants to Improve the Performance of *Tanacetum balsamita* L., a Medicinal Plant. Applied Sciences. 2023; 13(2):722. <https://doi.org/10.3390/app13020722>, [@2023](#) [Линк](#)
48. Sugier D., Sugier P., Jakubowicz-Gil J., Gawlik-Dziki U., Zajac A., Król B., Chmiel S., Kończak M., Pięt M., Paduch R. Nitrogen Fertilization and Solvents as Factors Modifying the Antioxidant and Anticancer Potential of *Arnica montana* L. Flower Head Extracts. Plants. 2023; 12(1):142. <https://doi.org/10.3390/plants12010142>, [@2023](#) [Линк](#)

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## 2014

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12. Zayova E, Petrova M, Dimitrova L, Vasilevska-Ivanova R, Stoeva D. Effect of different auxins on in vitro rooting of *Paulownia elongata* plants. Genetics and Plant Physiology, 4, 3-4, 155-162, 2014.

Цитира се е:

49. Kim TD, Kim NH, Park EJ, Lee NN. High-frequency regeneration of plants in vitro from seedling-derived apical bud explants of *Tilia mandshurica* Rupr. & Maxim. Journal of Plant Biotechnology 48, 1, 54-61, [@2021](#) [Линк](#)
50. Srinivasan P, Raja HD, Tamilvanan R. Efficient *in vitro* plant regeneration from leaf-derived callus and genetic fidelity assessment of an endemic medicinal plant *Ranunculus wallichianus* Wight & Arn. by using RAPD and ISSR markers. Plant Cell, Tissue and Organ Culture 147, 2, 413-420, [@2021](#) [Линк](#)
51. Salem, J., Hassanein, A., El-Wakil, D. A., & Loutfy, N.. Interaction between Growth Regulators Controls *In Vitro* Shoot Multiplication in *Paulownia* and Selection of NaCl-Tolerant Variants. Plants, 11(4), 498, 2022, [@2022](#) [Линк](#)

13. **Petrova, M.**, Zayova, E., Todorova, M., Stanilova, M. Enhancement of *Arnica montana* In-Vitro Shoot Multiplication and Sesquiterpene Lactones Production Using Temporary Immersion System. International Journal of Pharmaceutical Sciences and Research, 5 (12), 5170-5176.

Цитира се е:

52. De Carlo, A.; Tarraf, W.; Lambardi, M.; Benelli, C. Temporary Immersion System for Production of Biomass and Bioactive Compounds from Medicinal Plants. Agronomy 2021, 11, 2414. <https://doi.org/10.3390/agronomy11122414>, [@2021](#) [Линк](#)
53. Nieto-Trujillo, A.; Cruz-Sosa, F.; Luria-Pérez, R.; Gutiérrez-Rebolledo, G.A.; Román-Guerrero, A.; Burrola-Aguilar, C.; Zepeda-Gómez, C.; Estrada-Zúñiga, M.E. *Arnica montana* Cell Culture Establishment, and Assessment of Its Cytotoxic, Antibacterial, α-Amylase Inhibitor, and Antioxidant In Vitro Bioactivities. Plants 2021, 10, 2300. <https://doi.org/10.3390/plants10112300>, [@2021](#) [Линк](#)
54. Mirzabe, A. H., Hajiahmad, A., Fadavi, A., & Rafiee, S. Temporary immersion systems (TISs): A comprehensive review. Journal of Biotechnology, Volume 357, 20 September 2022, Pages 56-83, 2022, [@2022](#) [Линк](#)
55. Sugier D, Sugier P, Jakubowicz-Gil J, Gawlik-Dziki U, Zajac A, Król B, Chmiel S, Kończak M, Pięt M, Paduch R. Nitrogen Fertilization and Solvents as Factors Modifying the Antioxidant and Anticancer Potential of *Arnica montana* L. Flower Head Extracts. Plants. 2023; 12(1):142. <https://doi.org/10.3390/plants12010142>, [@2023](#)

14. Petrova, M., Zayova, E., Dincheva, I., Badjakov, I., Vlahova, M. Influence of carbon sources on growth and GC-MS based metabolite profiling of *Arnica montana* L. hairy roots. Turk J Biol, 39(3), 469-478, 2015.

Цитира се е:

56. Kentsop, R. A. D., Iobbi, V., Donadio, G., Ruffoni, B., De Tommasi, N., & Bisio, A. Abietane Diterpenoids from the Hairy Roots of *Salvia corrugata*. Molecules, 26(17), 5144., 2021., [@2021 Линк](#)
57. Khazaei-Poul, Y., Farhadi, S., Ghani, S., Ahmadizad, S. A., & Ranjbari, J. Monocyclic peptides: types, synthesis and applications. Current pharmaceutical biotechnology, 22(1), 123-135., 2021, [@2021 Линк](#)
58. Makowczyńska, J., Kalemba, D., & Skała, E. Establishment of *Codonopsis pilosula* (Franch.) Nannf. transformed roots, influence of the culture conditions on root growth and production of essential oil. Industrial Crops and Products, 165, 113446, 2021, [@2021 Линк](#)
59. Nieto-Trujillo, A.; Cruz-Sosa, F.; Luria-Pérez, R.; Gutiérrez-Rebolledo, G.A.; Román-Guerrero, A.; Burrola-Aguilar, C.; Zepeda-Gómez, C.; Estrada-Zúñiga, M.E. *Arnica montana* Cell Culture Establishment, and Assessment of Its Cytotoxic, Antibacterial,  $\alpha$ -Amylase Inhibitor, and Antioxidant *In Vitro* Bioactivities. Plants 2021, 10, 2300. <https://doi.org/10.3390/plants10112300>, [@2021 Линк](#)
60. Roy, A. Hairy Root Culture an Alternative for Bioactive Compound Production from Medicinal Plants. Current Pharmaceutical Biotechnology 22(1), 136-149, 2021., [@2021 Линк](#)
61. Stanojković, J., Todorović, S., Pećinar, I., Lević, S., Ćalić, S., & Janošević, D. Leaf glandular trichomes of micropropagated *Inula britannica*—Effect of sucrose on trichome density, distribution and chemical profile. Industrial Crops and Products, 160, 113101., [@2021 Линк](#)
62. Gyawali, N., Rayamajhi, A., Karki, D., Pokhrel, T., Adhikari, A. *Arnica montana* L.: Traditional Uses, Bioactive Chemical Constituents, and Pharmacological Activities.

In: Devkota, H.P., Aftab, T. (eds) Medicinal Plants of the Asteraceae Family. Springer, Singapore. [https://doi.org/10.1007/978-981-19-6080-2\\_4](https://doi.org/10.1007/978-981-19-6080-2_4), 2022, [@2022](#) [Линк](#)

63. Skała, E., Olszewska, M. A., Makowczyńska, J., & Kicel, A. Effect of Sucrose Concentration on *Rhaponticum carthamoides* (Willd.) Iljin Transformed Root Biomass, Caffeoylquinic Acid Derivative, and Flavonoid Production. International Journal of Molecular Sciences, 23(22), 13848., 2022., [@2022](#) [Линк](#)
64. Castro-Juárez, C. J., Luna-Suárez, S., de Fátima Rosas-Cárdenas, F., Villa-Ruano, N.. "Hernandulcin Production in Elicited Hairy Roots of *Phyla scaberrima*: Toward Sustainable Production of a Non-Caloric Sweetener with Nutraceutical Properties." Chemistry & Biodiversity, 21(3), e202302095, 2024, [@2024](#) [Линк](#)
65. Thanonkeo, S., Palee, T., Thanonkeo, P., Klanrit, P. "Influence of Culture Conditions on Growth and Daidzein and Genistein Production in Hairy Root Cultures of *Pueraria candolleana* var. *mirifica*." Horticulturae, 10(8), 788, 2024, [@2024](#) [Линк](#)
15. **Petrova, M.**, Nikolova, M., Dimitrova, L., Zayova, E. Micropropagation and evaluation of flavonoid content and antioxidant activity of *Salvia officinalis* L. Genetics and Plant Physiology, 5, 1, 48-60, 2015.

Цитира се въз:

66. Grzegorczyk-Karolak, I., Hnatuszko-Konka, K., Krzemińska, M., Olszewska, M. A., & Owczarek, A. Cytokinin-Based Tissue Cultures for Stable Medicinal Plant Production: Regeneration and Phytochemical Profiling of *Salvia bulleyana* Shoots. Biomolecules, 11(10), 1513, 2021, [@2021](#) [Линк](#)
67. Deepa, A. V., & Thomas, D. T. Tissue Culture Studies in Lamiaceae: A Review. Biotechnology and Crop Improvement, 181-211, 2022, [@2022](#) [Линк](#)
68. Martini, A. N., Vlachou, G., & Papafotiou, M. Effect of Explant Origin and Medium Plant Growth Regulators on In Vitro Shoot Proliferation and Rooting of *Salvia tomentosa*, a Native Sage of the Northeastern Mediterranean Basin. Agronomy, 12(8), 1889, 2022, [@2022](#) [Линк](#)
69. Rostami, F., Radjabian, T., & Abrishamchi, P. Enhancement of phenolic acids accumulation in *Salvia abrotanoides* (Kar.) Sytsma shoot cultures under elicitation

with nitric oxide. Plant Cell, Tissue and Organ Culture (PCTOC), 149(1), 441-453, 2022., @2022 [Линк](#)

70. Copetta A., Mussano P., Devi P., Lanteri A., Cassetti A., Mascarello C., Bisio A., Ruffoni B. In Vitro Micropropagation, Rooting and Acclimatization of Two Agastache Species (*A. aurantiaca* and *A. mexicana*). Horticulturae. 2023; 9(10):1065. <https://doi.org/10.3390/horticulturae9101065>, @2023 [Линк](#)
71. Grzegorczyk-Karolak, I., Krzemińska, M., Kiss, A. K., Owczarek-Januszkiewicz, A., Olszewska, M. A. Role of Phytohormones in Biomass and Polyphenol Accumulation in *Salvia bulleyana* *In Vitro* Culture. Biomolecules, 13(2), 227, 2023, @2023 [Линк](#)
72. Nanos C., Tsoulphra P., Kostas S., Hatzilazarou S., Michail I., Anastasiadi V., Pipinis E., Gklavakis E., Kanellis A.K., Nianiou-Obeidat I. Asexual Propagation of Greek *Salvia officinalis* L. Populations Selected for Ornamental Use. Horticulturae. 2023; 9(7):847. <https://doi.org/10.3390/horticulturae9070847>, @2023 [Линк](#)
73. Papafotiou, M., Vlachou, G., Martini, A. N. Investigation of the Effects of the Explant Type and Different Plant Growth Regulators on Micropropagation of Five Mediterranean *Salvia* spp. Native to Greece. Horticulturae, 9(1), 96, 2023, @2023 [Линк](#)
74. Sayed, S. S., Marzouk, M., Sokkar, N. Determination of Phenolics and Flavonoids with Antioxidant Effect of *Brunfelsia Pauciflora* (Cham. & Schtdl) Benth through *in Vitro* Propagated Cultures. Egyptian Journal of Chemistry, 66(4), 237-244, 2023., @2023 [Линк](#)
75. Jo, J.W., Yang, S.W., Lee, G.W., Kim, J.H., Kim, Y.J., Choi, Y.-K., Kim, K.J., Lee, H.-S., Bang, S.W., Kim, H.J. Effect of a Directional Electromagnetic Field on the Early Stages of Plant (*Raphanus sativus* and *Saccharum officinarum*) Growth. Horticulturae 2024, 10, 973. <https://doi.org/10.3390/horticulturae10090973>, 2024, @2024 [Линк](#)
76. Moshari-Nasirkandi, R., Alirezalu, A., Chamanabad, H. R. M., Amato, J., Alipour, H., Asghari, A., & Moshari-Nasirkandi, A. Screening of native wild *Salvia nemorosa* populations for chemical compositions, antioxidant activity and UHPLC-HRMS profiling. Scientific Reports, 14(1), 32064, 2024, @2024 [Линк](#)

---

2016

---

16. Zayova, E., **Petrova, M.**, Nikolova, M., Dimitrova, L. Effect of Medium Salt Strength on the Micropropagation, Phenolic Content and Antioxidant Activity of *Arnica montana* L., Threatened Plant Species. Bio Bulletin, 2, 1, 6-13, 2016.

Цитира се въз:

77. Abdalla N, El-Ramady H, Seliem MK, El-Mahrouk ME, Taha N, Bayoumi Y, Shalaby TA, Dobránszki J. An academic and technical overview on plant micropropagation challenges. Horticulturae, 8(8), 677, [@2022](#) [Линк](#)
78. Oseni, O. M., Nailwal, T. K., & Pande, V. Callus induction and multiple shoot proliferation from nodal explants of *Mansonia altissima*: confirmation of genetic stability using ISSR and RAPD markers. In Vitro Cellular & Developmental Biology-Plant, 58, 479–488, [@2022](#) [Линк](#)
17. Zayova, E, Nikolova, M, Dimitrova, L, **Petrova, M.** Comparative study of *in vitro*, *ex vitro* and *in vivo* propagated *Salvia hispanica* (CHIA) plants: morphometric analysis and antioxidant activity. AgroLife Scientific Journal, 5, 2, 166-173, 2016.

Цитира се въз:

79. Chiomento, J. L. T., De Nardi, F. S., Filippi, D., Trentin, T. D. S., Anzolin, A. P., Bertol, C. D., Nienow, A.A., Calvete, E. O. Mycorrhization of strawberry plantlets potentiates the synthesis of phytochemicals during ex vitro acclimatization. Acta Scientiarum. Agronomy, 44, 2022, [@2022](#) [Линк](#)
80. Martini, A. N., Vlachou, G., & Papafotiou, M. Effect of Explant Origin and Medium Plant Growth Regulators on In Vitro Shoot Proliferation and Rooting of *Salvia tomentosa*, a Native Sage of the Northeastern Mediterranean Basin. Agronomy, 12(8), 1889, 2022., [@2022](#) [Линк](#)
81. Motyka, S., Koc, K., Ekiert, H., Blicharska, E., Czarnek, K., & Szopa, A. The Current State of Knowledge on *Salvia hispanica* and *Salviae hispanicae semen* (Chia Seeds). Molecules, 27(4), 1207, 2022., [@2022](#) [Линк](#)
82. Wani, S., Kaloo, Z. A., Ganie, A. H., Shah, M. A., & Tali, B. A. Micropropagation and Arbuscular Mycorrhizae Assisted Growth in *Phlomis cashmeriana* Royle ex Benth.,

an endemic medicinal herb of Kashmir Himalaya. Journal of Herbs, Spices & Medicinal Plants, 28(3), 237-264, 2022., @2022 [Линк](#)

83. Copetta A., Mussano P., Devi P., Lanteri A., Cassetti A., Mascarello C., Bisio A., Ruffoni B. In Vitro Micropropagation, Rooting and Acclimatization of Two Agastache Species (*A. aurantiaca* and *A. mexicana*). Horticulturae. 2023; 9(10):1065. <https://doi.org/10.3390/horticulturae9101065>, @2023 [Линк](#)
84. Papafotiou, M., Vlachou, G., Martini, A. N. Investigation of the Effects of the Explant Type and Different Plant Growth Regulators on Micropropagation of Five Mediterranean *Salvia* spp. Native to Greece. Horticulturae, 9(1), 96, 2023., @2023 [Линк](#)
85. Bansal, M., Mujib, A., Bansal, Y., Dewir, Y. H., Mendler-Drienyovszki, N. An efficient In vitro shoot organogenesis and comparative GC-MS metabolite profiling of *Gaillardia pulchella* Foug. Horticulturae, 10(7), 728, 2024, @2024 [Линк](#)
86. Bansal, Y., Mujib, A., Mamgain, J., Kumar, S., Dewir, Y. H., Magyar-Tábori, K. Synthesis and Accumulation of Phytocompounds in Field-, Tissue-Culture Grown (Stress) Root Tissues and Simultaneous Defense Response Activity in *Glycyrrhiza glabra* L. Sustainability, 16(4), 1613, 2024, @2024 [Линк](#)
87. Bansal, Y., Mujib, A., Mamgain, J., Syeed, R., Mohsin, M., Nafees, A., Dewir, Y.H. , Mendler-Drienyovszki, N. Integrated GC-MS and UPLC-ESI-QTOF-MS based untargeted metabolomics analysis of in vitro raised tissues of *Digitalis purpurea* L. Frontiers in Plant Science, 15, 1433634, 2024, @2024 [Линк](#)
88. Fatima, T., Mujib, A., Bansal, Y., Dewir, Y. H., Mendler-Drienyovszki, N. Indirect Organogenesis of *Calendula officinalis* L. and Comparative Phytochemical Studies of Field-Grown and In Vitro-Regenerated Tissues. Agronomy, 14(8), 1743, 2024, @2024 [Линк](#)
89. García-Ramírez, Y. Temporary Immersion System for Biomass Production of *Salvia* spp.: A Mini-Review. Micropropagation Methods in Temporary Immersion Systems, 217-225, 2024, @2024 [Линк](#)
18. Todorova, M., Trendafilova, A., Vitkova, A., Petrova, M., Zayova, E., Antonova, D.. Developmental and Environmental Effects on Sesquiterpene Lactones in Cultivated *Arnica montana* L.. Chemistry & Biodiversity, 13(8), 976-981, 2016.

Цитира се въз:

90. Nieto-Trujillo, A.; Cruz-Sosa, F.; Luria-Pérez, R.; Gutiérrez-Rebolledo, G.A.; Román-Guerrero, A.; Burrola-Aguilar, C.; Zepeda-Gómez, C.; Estrada-Zúñiga, M.E. *Arnica montana* Cell Culture Establishment, and Assessment of Its Cytotoxic, Antibacterial,  $\alpha$ -Amylase Inhibitor, and Antioxidant In Vitro Bioactivities. *Plants* 2021, 10, 2300. <https://doi.org/10.3390/plants10112300>, @2021 [Линк](#)
91. Fusani, P., Aiello, N., Shachter, A., & Dudai, N. Volatile Composition Variability of *Arnica montana* Wild Populations of Trentino-Alto Adige, Italy, Determined by Headspace-Solid Phase Microextraction. *Chemistry & Biodiversity*, 19(1), e202100593, 2022, @2022 [Линк](#)
92. Greinwald, A., Hartmann, M., Heilmann, J., Heinrich, M., Luick, R., & Reif, A. Soil and Vegetation Drive Sesquiterpene Lactone Content and Profile in *Arnica montana* L. Flower Heads From Apuseni-Mountains, Romania. *Frontiers in plant science*, 13, 2022., @2022 [Линк](#)
93. Šadibolová, M., Juvonen, R. O., Auriola, S., & Boušová, I. *In vitro* metabolism of helenalin and its inhibitory effect on human cytochrome P450 activity. *Archives of Toxicology*, 96(3), 793-808, 2022., @2022 [Линк](#)
94. Parafiniuk, A., Kromer, K., Fleszar, M. G., Kreitschitz, A., Wiśniewski, J., Gamian, A. Localization of Sesquiterpene Lactones Biosynthesis in Flowers of Arnica Taxa. *Molecules*, 28(11), 4379., @2023 [Линк](#)
95. Schmidt, T. J. (2023). *Arnica montana* L.: Doesn't Origin Matter?. *Plants*, 12(20), 3532., @2023 [Линк](#)

---

2018

---

19. Zayova E, Geneva M, Stancheva I, Dimitrova L, **Petrova M**, Hristozkova M, Salamon I. Evaluation of the antioxidant potential of *in vitro* propagated hyssop (*Hyssopus officinalis* L.) with different plant growth regulators. *Medicinal Plants - International Journal of Phytomedicines and Related Industries*, 10(4), 295-304, 2018.

Цитира се въз:

96. Long, Y., Yang, Y., Pan, G., & Shen, Y. New insights into tissue culture plant-regeneration mechanisms. *Frontiers in Plant Science*, 13, 2022., [@2022](#) [Линк](#)
97. Chokheli VA, Bakulin SD, Ermolaeva OY, Kozlovsky BL, Dmitriev PA, Stepanenko VV, Kornienko IV, Bushkova AA, Rajput VD, Varduny TV. Investigation of Growth Factors and Mathematical Modeling of Nutrient Media for the Shoots Multiplication In Vitro of Rare Plants of the Rostov Region. *Horticulturae*. 9(1):60., [@2023](#) [Линк](#)
98. Pal, D., Kumar, M., Yadav, M. K., Chauhan, C., Kumar, A., Sirohi, U., Rakesh Sharma, V. & Chaudhary, V. Effect of various chemical solutions on micropropagation efficiency, antioxidant activities and clonal fidelity analysis of in vitro developed plantlets in pomegranate (*Punica granatum* L.) using SSR markers. *Vegetos*, 1-10., 2024, [@2024](#) [Линк](#)
99. Popova EA, Pungin AV, Pantyukhina AP, Krol OV. Evaluating secondary metabolites and antioxidant activity of in vitro callus and micro-plant extracts of *Hyssopus officinalis* L. *Food Processing: Techniques and Technology*. 54(4): 658–674. (In Russ.), [@2024](#) [Линк](#)
20. Zayova E, Stancheva I, Geneva M, Hristozkova M, Dimitrova L, **Petrova M**, Sichanova M, Salamon I, Mudroneckova S. Arbuscular mycorrhizal fungi enhance antioxidant capacity of in vitro propagated garden thyme (*Thymus vulgaris* L.). *Symbiosis*, 74, 177-187, 2018.

Цитира се в:

100. Biel C, Camprubí A, Lovato P, Calvet C, . On-farm reduced irrigation and fertilizer doses, and arbuscular mycorrhizal fungal inoculation improve water productivity in tomato production, *Scientia Horticulturae* 288 (2021) 110337, [@2021](#) [Линк](#)
101. Rajsza A, Wojtuń B, Samecka-Cymerman A, Wąsowicz P, Mróz L, Rudecki A, Kempers AJ, . Metals in *Calluna vulgaris*, *Empetrum nigrum*, *Festuca vivipara* and *Thymus praecox* ssp. *arcticus* in the geothermal areas of Iceland. *Environ Sci Pollut Res* 28, 67224–67233, [@2021](#) [Линк](#)
102. Sun D, Shang X, Cao H, Lee S-J, Wang L, Gan Y, Feng S. Physio-Biochemical Mechanisms of Arbuscular Mycorrhizal Fungi Enhancing Plant Resistance to Abiotic Stress. *Agriculture*. 14(12):2361, [@2024](#) [Линк](#)

103. Nordine, A. Trends in plant tissue culture, production, and secondary metabolites enhancement of medicinal plants: a case study of thyme. *Planta*, 261(4), 84, [@2025](#) [Линк](#)
104. Sahoo, M., Kullu, B. *Claroideoglomus claroideum* inoculation improves biohardening of micropropagated G9 banana by modulating antioxidative response, nutrient uptake and photosynthetic efficiency. *Discov. Plants* 2, 110 (2025). <https://doi.org/10.1007/s44372-025-00194-4>, [@2025](#)
- 

## 2019

---

21. **Petrova M**, Zayova E, Dimitrova L, Geneva M, Miladinova-Georgieva K. Micropropagation studies and antioxidant analysis of the endangered plants of Bulgarian yellow gentian (*Gentiana lutea* L.). *Acta Scientiarum Polonorum Hortorum Cultus*, 18, 3, 71-78, 2019.

Цитира се в:

105. Ponticelli M, Lela L, Moles M, Mangieri C, Bisaccia D, Faraone I, Falabella R, Milella L. "The healing bitterness of *Gentiana lutea* L., phytochemistry and biological activities: A systematic review". *Phytochemistry*, Volume 206, 2023, 113518., [@2022](#) [Линк](#)
106. Demirci, T., Albayrak, I., Deveci, H., Asci, Ö. A., & Baydar, N. G. "Development of effective micropagation protocols for endangered *Gentiana lutea* and *Gentiana boissieri* (endemic to Türkiye)." *Israel Journal of Plant Sciences*, 71(3-4), 127-136, 2024, [@2024](#) [Линк](#)
- 

## 2020

---

22. Todorova D, Katerova Z, Dimitrova R, **Petrova M**, Hristozkova M, Sergiev I. Exogenous spermine application increases quantity of rosmarinic acid and carnosic acid in salt-treated *Salvia officinalis* L. plants in pot experiments. *Compt. Rend. Acad. Bulg. Sci.*, 73 (6), 800-808, 2020

Цитира се в:

- 107.** Gholamnia A, Arani AM, Sodaeizadeh H, Esfahani ST, Ghasemi S (2022) Expression profiling of rosmarinic acid biosynthetic genes and some physiological responses from *Mentha piperita* L. under salinity and heat stress. *Physiol Mol Biol Plants* 28, 545–557. <https://doi.org/10.1007/s12298-022-01159-7>, @2022 [Линк](#)
- 108.** Shahtousi S, Talaee L . The effect of spermine on *Tetranychus urticae-Cucumis sativus* interaction. *BMC Plant Biology*, 23(1), 575 <https://doi.org/10.1186/s12870-023-04573-5>, 2023, @2023 [Линк](#)
- 

## 2021

---

- 23.** **Petrova M**, Zayova E, Geneva M, Dimitrova L, Vitkova A, Stanilova M. Multiplication and conservation of threatened medicinal plant *Arnica montana* L. by in vitro techniques. *Agriculturae Conspectus Scientificus*, 86, 1, 57-65, 2021.

Цитира се е:

- 109.** Riahi, L., Chograni, H., Ben Rejeb, F., Ben Romdhane, M., Masmoudi, A. S., & Cherif, A. Efficient *in vitro* regeneration of the endangered species *Artemisia arborescens* L. through direct organogenesis and impact on secondary metabolites production. *Horticulture, Environment, and Biotechnology*, 63, 439–450, @2022 [Линк](#)
- 110.** Nazari, M., Kordrostami, M., Ghasemi-Soloklui, A.A. Conservation of Medicinal Plants by Tissue Culture Techniques. In: Jha, S., Halder, M. (eds) Medicinal Plants: Biodiversity, Biotechnology and Conservation. Sustainable Development and Biodiversity, Vol. 33, Springer, Singapore, @2023 [Линк](#)
- 111.** Armijos-González, R., Ramón, P., Cueva-Agila, A. "Cinchona officinalis L. ex situ conservation by *in vitro* slow growth and cryopreservation techniques." *Plant Cell, Tissue and Organ Culture (PCTOC)*, 158(1), 6., @2024 [Линк](#)
- 112.** Linjikao, J., Inthima, P., Limmongkon, A., & Kongbangkerd, A. "Mannitol and sorbitol concentration optimization for effective *Epipactis flava* Seidenf. *in vitro* slow growth storage." In *Vitro Cellular & Developmental Biology-Plant*, 60, 496–507., @2024 [Линк](#)
- 24.** **Petrova M.**, Nikolova M., Dimitrova M., Dimitrova L. Assessment of the effect of plant growth regulators on *in vitro* micropropagation and metabolic profiles of *Melissa officinalis*

l. (lemon balm). Journal of Microbiology, Biotechnology and Food Sciences, 11(3), e4077-e4077, 2021.

Цитира се в:

113. Deepa, A. V., & Thomas, D. T. Tissue Culture Studies in Lamiaceae: A Review. Biotechnology and Crop Improvement, 181-211, 2022., [@2022](#) [Линк](#)
  114. Yegorova, N. A., Yakimova, O. V., & Belova, I. V. "Long-Term Passage and Characteristics of *Melissa officinalis* L. Callus Cell Cultures." Russian Journal of Plant Physiology, 71(3), 101, 2024, [@2024](#) [Линк](#)
- 

## 2022

---

25. Sichanova M, Geneva M, **Petrova M**, Miladinova-Georgieva K, Kirova E, Nedev T, Tsekova D, Iwanov I, Dochev K, Ivanova V, Trendafilova T. Improvement of *Stevia rebaudiana* Bertoni *in vitro* propagation and steviol glycoside content using aminoacid silver nanofibers. Plants, Plants, 11(19), 2468, 2022.

Цитира се в:

115. Coman V, Scurtu V, Coman C, Clapa D, Iancu S, Leopold N, Leopold L, Effects of polystyrene nanoplastics exposure on *in vitro*-grown *Stevia rebaudiana* plants, 197, 107634, Plant Physiology and Biochemistry, 197, 107634, [@2023](#) [Линк](#)
116. Ptak, A , Szewczyk, A, Simlat, M, Błażejczak, A & Warchał, M. "Meta-Topolin-induced mass shoot multiplication and biosynthesis of valuable secondary metabolites in *Stevia rebaudiana* Bertoni bioreactor culture". Sci Rep 13, 15520, 2023, [@2023](#) [Линк](#)
117. Sharma S, Gupta S, Jain R, Kothari S L, Kachhwaha S, SiO<sub>2</sub> nanoparticles as elicitor for increased rebaudioside-A in *Stevia rebaudiana* micropagated in solid and liquid cultures: a comparative study, Plant Cell, Tissue and Organ Culture (PCTOC), 155, 541–552, [@2023](#) [Линк](#)
118. Sharma S, Gupta S, Kumari D, Lal Kothari S, Jain R, Kachhwaha S, Exploring plant tissue culture and steviol glycosides production in *Stevia rebaudiana* (Bert.) Bertoni: A Review, Agriculture 13(2):475, [@2023](#) [Линк](#)

119. Bouaaza, G Chetto, O Beniken, L Dziri, L M M'hamed, M Benkirane, R Benyahia, H. "Effect of gelling agent and plant growth regulators on in vitro shooting and rooting of *Stevia rebaudiana* Bertoni". Advances in Agriculture. Article ID 5131187, @2024 [Линк](#)
120. Jadid N, Anggraeni S, Ramadani MRN, Arieny M, Mas'ud F, In vitro propagation of Indonesian stevia (*Stevia rebaudiana*) genotype using axenic nodal segments. BMC Res Notes 17, 45, @2024 [Линк](#)
121. Mercado-Díaz de León L., Garcidueñas-Piña C., Pérez-Molphe-Balch E., Loera-Muro A., Morales-Domínguez J. F. Effect of Bv AgNP on growth, development, and glyoxalase gene expression analysis in *Mammillaria bombycina* and *Selenicereus undatus*. Molecular Biology Reports, 51(1), 681, 2024, @2024 [Линк](#)
122. Subrahmanyewari T, Gantait S, Sarkar R, Kamble SN, Singh S, Bhattacharyya S Polyamines- and growth inducers-mediated enhanced mono-phasic in vitro regeneration of sugar leaf plant (*Stevia rebaudiana* Bert.) in liquid medium, South African Journal of Botany, 173, 34-45, @2024 [Линк](#)
123. Karki R, Ojha P, Maharjan S, Manandhar U, Maharjan S, Optimization of the germination time of proso and foxtail millets to enhance the bioactive properties, antioxidant activity, and enzymatic power and reduce antinutritional factor, Curr Res in Food Sci, 10, 100987, @2025 [Линк](#)
124. Manohar KA, Sivasankarreddy K, Shukla G, Chakravarty S, Roy B, Chakravarty S, Optimization of an Enhanced Micropropagation Protocol of *Stevia rebaudiana* for Mass Production in an Indian Sub-humid Region. Sugar Tech, <https://doi.org/10.1007/s12355-025-01542-0>, @2025 [Линк](#)
125. Pal D, Kumar M, Yadav MK, Chauhan C, Kumar A, Sirohi U, Sharma V, Chaudhary V Effect of various chemical solutions on micropropagation efficiency, antioxidant activities and clonal fidelity analysis of *in vitro* developed plantlets in pomegranate (*Punica granatum* L.) using SSR markers. Vegetos, 38, 796–805 (2025), @2025 [Линк](#)
26. Kirova E, Geneva M, **Petrova M**, Miladinova-Georgieva K, Sichanova M. Employment of nanoparticles for improvement of plant growth and development. Botanica, 28, 2, 113-132, 2022.

Цитира се въз:

126. Dönmez, D., Isak, M. A., İzgü, T., & Şimşek, Ö. "Green Horizons: Navigating the Future of Agriculture through Sustainable Practices." *Sustainability*, 16(8), 3505, **@2024** [Линк](#)
127. Martínez-Chávez LA, Hernández-Ramírez MY, Feregrino-Pérez AA, Esquivel Escalante K. Cutting-edge strategies to enhance bioactive compound production in plants: Potential value of integration of elicitation, metabolic engineering, and green nanotechnology. *Agronomy*. 14(12):2822, **@2024** [Линк](#)
128. Yousaf, M., Munir, R. M., Iqbal, T., Afsheen, S., Khan, M. I., Wali, H., Rizvi H. I., Pham Ph. V. , AlObaid A. A. , Warad I. , Rafique, S. Exploring innovative antibacterial properties of porous ALT (Al<sub>2</sub>O<sub>3</sub>/TiO<sub>2</sub>) composite. *Materials Chemistry and Physics*, 325, 129736, 2024, **@2024** [Линк](#)
27. Stanilova M, Traykova B, Vladimirov V, **Petrova M**, Semerdjieva I, Yankova-Tsvetkova E. In vitro micropropagation of *Helichrysum arenarium* (Asteraceae) as a tool for introducing the species in agriculture. *Comptes rendus de l'Académie bulgare des Sciences*, 75, 10, 1454-1461, 2022.

Цитира се въз:

129. Cuce, M., Inceer, H. Micropropagation and reintroduction of the endemic *Tripleurospermum ziganaense* (Asteraceae) to its natural habitat. *In Vitro Cell.Dev.Biol.-Plant* 60, 646–658 (2024). <https://doi.org/10.1007/s11627-024-10457-6>, **@2024** [Линк](#)
28. Yankova-Tsvetkova E, **Petrova M**, Grigorova I, Traykova B, Stanilova M. The establishment of an ex situ collection of *Primula veris* in Bulgaria. *Plants*, 11(22), 3018, 2022.

Цитира се въз:

130. Graikou, K., Mpishinioti, A., Tsafantakis, N., Maloupa, E., Grigoriadou, K., & Chinou, I. Comparative phytochemical analyses of flowers from *Primula veris* subsp. *veris* growing wild and from ex situ cultivation in Greece. *Foods*, 12(13), 2623, 2023, **@2023** [Линк](#)

131. Sarropoulou, V., Sarrou, E., Angeli, A., Martens, S., Maloupa, E., & Grigoriadou, K. Developing an *in vitro* elicitation strategy for specialized secondary metabolites production in adventitious root cultures of *Primula veris* subsp. *veris*. Industrial Crops and Products, 197, 116618, 2023., [@2023](#) [Линк](#)
29. Geneva M, Stancheva I, Kirova E, **Petrova M**, Hendawy S, Zayova E. Assessment of antioxidant activity of in vitro obtained plant of *Coleus forskohlii* Briq. Journal of Microbiology, Biotechnology and Food Sciences, 11(4), e3840-e3840, 2022

Цитира се е:

132. Badhepuri M, Manokari M, Cokul Raj M, Jogam P, Dey A, Faisal M, Alatar A, Joshee N, Singisala N, Shekhawat M, Meta-Topolin enhanced direct shoot organogenesis and regeneration from leaf explants of *Coleus forskohlii* (Willd.) Briq, Industrial Crops and Products, 197, 116584, <https://doi.org/10.1016/j.indcrop.2023.116584>, [@2023](#) [Линк](#)
133. El-Fakharany EM, El-Maradny YA, Ashry M, Abdel-Wahhab KG, Shabana ME, El-Gendi H. Green synthesis, characterization, anti-SARS-CoV-2 entry, and replication of lactoferrin-coated zinc nanoparticles with halting lung fibrosis induced in adult male albino rats. Sci Rep 13, 15921, [@2023](#) [Линк](#)
30. Abou Obeid Y, Zehirov G, Again N, **Petrova M**, Haddad R, Karam F, Shaban N, Vassilevska-Ivanova R. Effects of inorganic and foliar fertilizers on antioxidant capacity and flower yield of saffron (*Crocus sativus* L.). Acta Agrobotanica, 75, 1, 754.

Цитира се е:

134. Alhasan, A. S. (2023). Effect of npk fertilizer on stigma and corm production in saffron (*Crocus sativus* L.). Int. J. Agricult. Stat. Sci. Vol, 19(1), 311-316, [@2023](#) [Линк](#)
135. Krzyminska-Brodka, A., et al. "The longevity of cut *Polygonatum multiflorum* (L.) All. shoots depending on postharvest handling." Acta Agrobotanica 76 (2023)., [@2023](#) [Линк](#)

31. Miladinova-Georgieva K, Geneva M, Stancheva I, **Petrova M**, Sichanova M, Kirova E. Effects of different elicitors on micropropagation, biomass and secondary metabolite production of *Stevia rebaudiana* Bertoni – a review. Plants, 12, 1, 153, 2023.

Цитира се е:

136. Ahmad, M. A., Chaudhary, S., Deng, X., Cheema, M., & Javed, R. "Nano-stevia interaction: Past, present, and future. " Plant Physiology and Biochemistry, 201, 107807, 2023, [@2023 Линк](#)
137. Clapa, D., Radomir, A.M., Petelică, A.G., Hărța, M. "Evaluation of biomass production of *Stevia rebaudiana* Bertoni using classical in vitro culture and temporary immersion bioreactor system" Scientific Papers. Series B, Horticulture. Vol. LXVII, No. 1, 558-565, 2023, [@2023 Линк](#)
138. Coman, V., Scurtu, V. F., Coman, C., Clapa, D., Iancu, Ş. D., Leopold, N., & Leopold, L. F. "Effects of polystyrene nanoplastics exposure on in vitro-grown *Stevia rebaudiana* plants." Plant Physiology and Biochemistry, 197, 107634, 2023, [@2023 Линк](#)
139. Jeyasri, R., Muthuramalingam, P., Karthick, K., Shin, H., Choi, S. H., & Ramesh, M. "Methyl jasmonate and salicylic acid as powerful elicitors for enhancing the production of secondary metabolites in medicinal plants: an updated review. "Plant Cell, Tissue and Organ Culture (PCTOC), 153(3), 447-458, 2023, [@2023 Линк](#)
140. Kanthaliya, B., Joshi, A., Arora, J., Alqahtani, M. D., & Abd\_Allah, E. F. "Effect of biotic elicitors on the growth, antioxidant activity and metabolites accumulation in *in vitro* propagated shoots of *Pueraria tuberosa*." Plants, 12(6), 1300, 2023, [@2023 Линк](#)
141. Ptak, A., Szewczyk, A., Simlat, M., Błażejczak, A., & Warchoł, M. "Meta-Topolin-induced mass shoot multiplication and biosynthesis of valuable secondary metabolites in *Stevia rebaudiana* Bertoni bioreactor culture. " Scientific Reports, 13(1), 15520., [@2023 Линк](#)
142. Wu M., Chen J., Tang W., Jiang Y., Hu Z., Xu D., Hou K., Chen Y., Wu W. "Genome-Wide Identification and Expression Analysis of bZIP Family Genes in *Stevia rebaudiana*" Genes 14(10):1918, CC BY 4.0, [@2023 Линк](#)

143. Altun, H., Orcan, P. "Responses to exogenous elicitor treatment in lead-stressed *Oryza sativa* L." BMC Plant Biology, 24(1), 897, @2024 [Линк](#)
144. Andiç B, Orcan P, *Salvia nemorosa* L. responses to silver nanoparticle and methyl jasmonate elicitors under salt stressjasmonate elicitors under salt stress, Turk J Bot, 48, 308-320, @2024 [Линк](#)
145. Biswas, P., Kumari, A., Modi, A., Kumar, N. "Improvement and regulation of steviol glycoside biosynthesis in *Stevia rebaudiana* Bertoni." Gene, 891, 147809, @2024 [Линк](#)
146. Medeiros APR, Leite JJF, de Assis RMA, Rocha JPM, Bertolucci SKV, Pinto JEBCP "Application of natural elicitors to promote growth, photosynthetic pigments, and the content and composition of essential oil in *Melissa officinalis* L." Industrial Crops and Products, 208, 117885, @2024 [Линк](#)
147. Mehdizadeh, L., Moghaddam, M., Ganjeali, A., & Mahmoodi Sourestani, M. "Changes in growth, essential oil composition and biochemical traits of peppermint in response to coapplication of zinc and methyl jasmonate in soilless culture." Journal of Plant Nutrition, 1-21, @2024 [Линк](#)
148. Papaefthimiou, M., Kontou, P. I., Bagos, P. G., Braliou, G. G. " Integration of Antioxidant Activity Assays Data of Stevia Leaf Extracts: A Systematic Review and Meta-Analysis." Antioxidants, 13(6), 692, @2024 [Линк](#)
149. Ptak A, Szewczyk A, Simlat M, Pawłowska B, Warchoł M "LED light improves shoot multiplication, steviol glycosides and phenolic compounds biosynthesis in *Stevia rebaudiana* Bertoni in vitro culture." Scientific Reports, 14(1), 30860, @2024 [Линк](#)
150. Sale S, Subramaniam S, Mad'Atari MF. "Trends in the tissue culture techniques and the synthesis of bioactive compounds in *Eurycoma longifolia* Jack—current status and future perspectives." Plants, 13(1), 107, @2024 [Линк](#)
151. Singh, S., Uddin, M., Singh, S., Ahmed, K. B. M., Bhat, U. H., Chishti, A. S., Chauhan, A.& Khan, M. M. A. "Radiolytically Depolymerized Low Molecular-Weight Chitosan (ICH) and Sodium Alginate (ISA) Improve Growth Attributes, Physiological Performance and the Production of Steviol Glycosides (SGs) of *S. rebaudiana* (Bertoni)." Journal of Polymers and the Environment, 32, 3258–3284, 2024, @2024 [Линк](#)

152. Śniegowska, J., Biesiada, A., Gasiński, A. "Influence of the Nitrogen Fertilization on the Yield, Biometric Characteristics and Chemical Composition of *Stevia rebaudiana* Bertoni Grown in Poland. " *Molecules*, 29(8), 1865, @2024 [Линк](#)
153. Wang, L., Zhu, T. "Combined transcriptomic and metabolomic analysis of the mechanism by which *Bacillus velezensis* induces resistance to anthracnose in walnut. " *Frontiers in Microbiology*, 15, 1420922, @2024 [Линк](#)
154. Khyahrii A. S., Shetty S., Kushalan S., Hegde S. Enhanced Curculigoside and Phenolic Compounds Elevate Antioxidant Activity in *C. orchoides* Gaertn In Vitro Under Exogenous Augmentation with Elicitors Salicylic Acid (SA) and Polyethylene Glycol (PEG) 6000. *Journal of Health and Allied Sciences NU*, 15(01), 125-131, 2025, @2025 [Линк](#)
155. Partap M., Kumar A., Kumar P., Kumar D., Warghat A. R. Elicitors Mediated Enhancement of Picrosides and Their Pathway Precursors in Dedifferentiated Cell Suspension Culture of *Picrorhiza kurroa* Royle ex Benth. *Journal of Plant Growth Regulation*, 44(1), 247-265, 2025, @2025 [Линк](#)
32. **Petrova M**, Dimitrova L, Dimitrova M, Denev P, Teneva D, Georgiieva A, Petkova-Kirova P, Lazarova M, Tasheva K. Antitumor and antioxidant activities of in vitro cultivated and wild-growing *Clinopodium vulgare* L. plants. *Plants*, 12, 8, 1591.

Цитира се в:

156. Vlachou G., Papafotiou, M., Daferera D.J., Tarantilis P.A. "Yield and Composition of the Essential Oil of *Clinopodium nepeta* subsp. *spruneri* as Affected by Harvest Season and Cultivation Method, i.e., Outdoor, Greenhouse and In Vitro Culture." *Plants* 2023, 12, 4098. <https://doi.org/10.3390/plants12244098>, @2023 [Линк](#)
157. Ahmed, K. A. A., Jabbar, A. A., M. Raouf, M. M. H., Al-Qaaneh, A. M., Mothana, R. A., Alanzi, A. R., Abdullah, F. O., Abdulla, M. A. , Hasson, S. , & Zainel, M. A. "Wood calamint ameliorates ethanol-induced stomach injury in rats by augmentation of hsp/bax and inflammatory mechanisms. " *Journal of Molecular Histology*, 55, 567–579, @2024 [Линк](#)
158. Konstantinou, E. K., Gioxari, A., Dimitriou, M., Panoutsopoulos, G. I., & Panagiotopoulos, A. A. "Molecular Pathways of Genistein Activity in Breast Cancer Cells." *International Journal of Molecular Sciences*, 25(10), 5556., @2024 [Линк](#)

159. Konstantinou, E. K., Panagiotopoulos, A. A., Argyri, K., Panoutsopoulos, G. I., Dimitriou, M., & Gioxari, A. Molecular pathways of rosmarinic acid anticancer activity in triple-negative breast cancer cells: a literature review. *Nutrients*, 16(1), 2., 2024, [@2024](#) [Линк](#)
160. Ogwu, M.C. "Systematics, Taxonomy, and Sustainable Management of Some Critical Vascular Plant Groups in Central Apennines, Italy". In: Al-Khayri, J.M., Jain, S.M., Penna, S. (eds) Sustainable Utilization and Conservation of Plant Genetic Diversity. Sustainable Development and Biodiversity, vol 35. Springer, Singapore. [https://doi.org/10.1007/978-981-99-5245-8\\_17](https://doi.org/10.1007/978-981-99-5245-8_17), [@2024](#) [Линк](#)
161. Qureshi, K. A., Parvez, A., Khan, M. M. U., Aspatwar, A., Atiya, A., Elhassan, G. O., Khan, R. A., Erattil Ahammed S. Y., Khan, W. U. & Jaremko, M. "Exploring nature's hidden treasure: Unraveling the untapped phytochemical and pharmacological potentials of *Clinopodium vulgare* L.—A hidden gem in the Lamiaceae family." *Heliyon*, Volume 10, Issue 2, e24781, [@2024](#) [Линк](#)
162. H Al-Qaisi, T. S., Raouf M. M. H. M., AbdulSamad Ismail P., Mothana R. A., Hawwal M. F., Hassan R. R., Abdulla M.A., Saleh M. I. , Awad M. (2025). Persimmon (*Diospyros kaki* L.) leaves accelerates skin tissue regeneration in excisional wound model: possible molecular mechanisms. *Journal of Molecular Histology*, 56, 73., [@2025](#) [Линк](#)
163. Li S, Zhang L, Zhang W, Chen H, Hong M, Xia J, Zhang W, Luan X, Zheng G, Lu D. Identifying traditional Chinese medicine combinations for breast cancer treatment based on transcriptional regulation and chemical structure. *Chin Med.*;20(1):23, 2025, [@2025](#) [Линк](#)
33. Sichanova M, Geneva M, **Petrova M**, Miladinova-Georgieva K, Kirova E, Nedev T, Tsekova D, Ivanova V, Trendafilova A. Influence of the abiotic elicitors Ag salts of aspartic acid derivatives, self-organized in nanofibers with monomeric and dimeric molecular structures, on the antioxidant activity and stevioside content in micropropagated *Stevia rebaudiana* Bert.. *Plants*, 12(20), 3574, 2023.

Цитира се е:

164. Vu, D. T., Kletthagen, M. C., Ellevoll, E. O., Falch, E., & Jensen, I. J. "Simulated Digestion of Red Sea Cucumber (*Parastichopus tremulus*): A Study of Protein Quality and Antioxidant Activity. " *Applied Sciences*, 14(8), 3267, 2024, [@2024](#) [Линк](#)

34. Tasheva K, Georgieva A, Denev P, Dimitrova L, Dimitrova M, Misheva S, Petkova-Kirova P, Lazarova M, **Petrova M.** Antioxidant and antitumor potential of micropropagated Balkan endemic *Sideritis scardica* Griseb. Plants, 12, 3924, 2023.

Цитира се е:

165. Wang R, Li Y, Ji J, Kong L, Huang Y, Liu Z, Lu L. The emerging role of herbal medicines in cancer by interfering with post-translational modification. *Antioxidants & Redox Signaling*, 42, 1-3, <https://doi.org/10.1089/ars.2023.0418>, @2024 [Линк](#)

166. Yücer R., Schröder A., Topçu G., Efferth T. Identification of anti-inflammatory and anti-cancer compounds targeting the NF-κB-NLRP3 inflammasome pathway from a phytochemical library of the *Sideritis* genus. *Journal of Ethnopharmacology*, Vol. 338, Part 2, 10 February 2025, 119074, @2025 [Линк](#)

---

2024

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35. **Petrova M**, Miladinova-Georgieva K, Geneva M. Influence of abiotic and biotic elicitors on organogenesis, biomass accumulation, and production of key secondary metabolites in Asteraceae plants. *International Journal of Molecular Science*, 25(8), 4197.

Цитира се е:

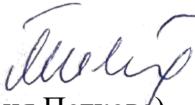
167. Benabderrahmane W, Fadel H, Sekhara I, Mennai I, Kadi IE, Helal M, Sami R, Abo-Dief HM, Bedaiwi RI, Alanazi MA, Al-Harthi HF, Kadi RH, Abushal SA, Albishi TS, Qumsanil AT, Qaril SH. GC-MS analysis, phytochemical composition of *Hertia cheirifolia* L. essential oil with pharmacological assessments: antioxidant, antibacterial, and antifungal activities. *RSC Adv.* 14, 22548-22559, @2024 [Линк](#)

168. Gergov E, Petrova P, Arsov A, Ignatova I, Tsigoriyna L, Armenova N, Petrov K. Inactivation of sacB gene allows higher 2, 3-butanediol production by *Bacillus licheniformis* from Inulin. *International Journal of Molecular Sciences* 25, 22, 11983, @2024 [Линк](#)

169. Patil J G, Nikam TD, Shinde RA, Ahire ML. Effect of abiotic and biotic elicitors on vincristine accumulation in endosperm derived in vitro cultures in *Catharanthus roseus* (L.) G. Don. *Discover Plants* 1, 1, 8, @2024 [Линк](#)

170. Ramakrishnan DK, Jauernegger F, Hoefle D, Berg C, Berg G, Abdelfattah A. Unravelling the microbiome of wild flowering plants: a comparative study of leaves and flowers in alpine ecosystems. *BMC Microbiology* 24, 1, 417, [@2024](#) [Линк](#)
171. Billowria P, Hussain S, Kapoor N, Mahajan R. Enhanced colchicine production in hairy root cultures of *Gloriosa superba* L. using microbial elicitors. *Indian Journal of Microbiology*, 1-14, [@2025](#) [Линк](#)
172. Ferreira, M.J., Garcia-Cardesín, E., Sierra-Garcia, I.N. , Pinto D.C.G.A., Cremades, J., Silva, H., Cunha, A. PGPB-driven bioenrichment and metabolic modulation of *Salicornia europaea* under marine Aquaponic conditions. *World J Microbiol Biotechnol* 41, 124 , 2025, [@2025](#) [Линк](#)
173. Parthasarathy SP, Elayaraja D, Archana A, Vignesh S, Sahayarayan JJ, Alagumanian S, Manickavasagam M, Unravelling the influence of precursors and polyamines on L-Dopa bio-fabrication in elicited suspension cells of *Hybanthus enneaspermus* (L.) F. Muell.. *Plant Cell Tiss Organ Cult* 161, 3, [@2025](#) [Линк](#)
174. Singh G, Passari AK, Kumar NS, Kumar B, Nayak SC, Ram H, Singh BP. UPLC-ESI MS/MS-and GC-MS-based altitudinal variations in the bioactive potential of *Mikania micrantha* and *Ageratum houstonianum*. *Applied Biochemistry and Biotechnology*, 197(1), 335-354, [@2025](#) [Линк](#)

Април 2025

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